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Bhardwaj et al.

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(54) **SUPEROXIDE DISMUTASE (SOD) GENE AND A METHOD OF IDENTIFYING AND CLONING THEREOF**

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(65) **Prior Publication Data**

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Related U.S. Application Data

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(30) **Foreign Application Priority Data**

Mar. 31, 2006 (IN) 928/DEL/2006

(51) **Int. Cl.**
C12N 9/02 (2006.01)
C12N 1/20 (2006.01)
C12N 15/00 (2006.01)
C12Q 1/68 (2006.01)
C07H 21/04 (2006.01)
C07H 21/02 (2006.01)

(52) **U.S. Cl.** **435/189**; 435/6; 435/252.3; 435/320.1; 536/23.1; 536/23.2

(58) **Field of Classification Search** None
See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

6,485,950 B1 11/2002 Kumar et al.
7,037,697 B2 5/2006 Kumar et al.

Primary Examiner—Yong D Pak

(74) *Attorney, Agent, or Firm*—Edwards Angell Palmer & Dodge LLP; Jeffrey D. Hsi; Elizabeth Spar

(57) **ABSTRACT**

The present invention provides a superoxide dismutase gene from *Potentilla atrosanguinea*, a construct containing the gene coding for superoxide dismutase and transformed *E. coli* producing the SOD protein.

5 Claims, 28 Drawing Sheets

Fig. 1

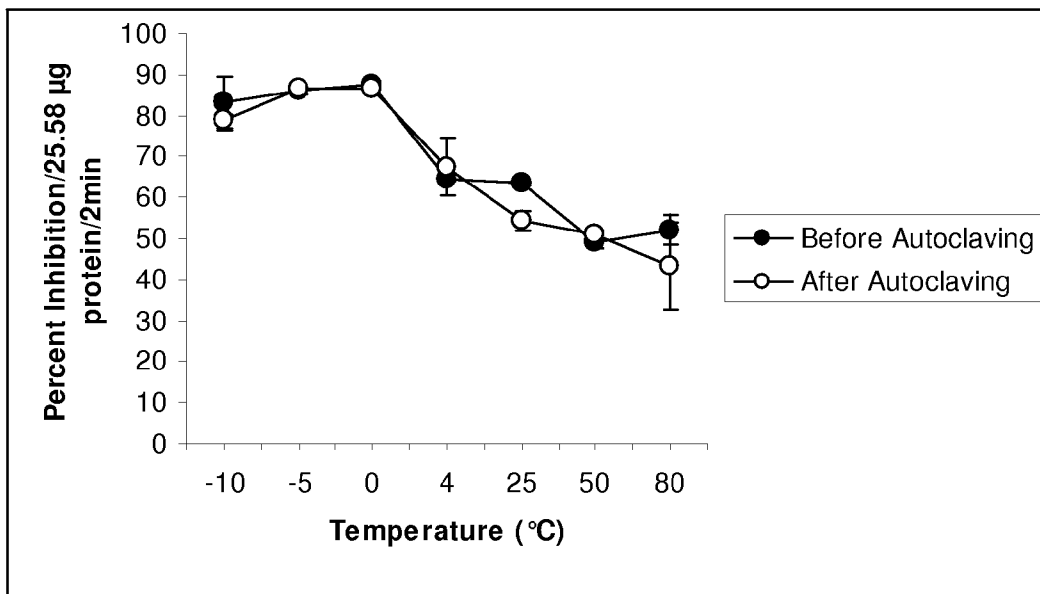


Fig. 2. Comparison of the nucleotide sequence of the *Potentilla* Cu/Zn SOD with sequences from other plant species. Regions of complete homology are indicated with asterisks.

Malus	ATGGTGAAGGGTGTGGCTGTTCTCGGCTCCAGTGAGGGCGTTAAAGGAACCATCAGCTTT
Potentilla	ATGGCAAAGGGCGTGTGCTACTTACTGCTCCAGTGAGGGTGTGGCTGGAACATCCTCTTT
Populus	ATGGTGAAGGGTGTAGCTGTTCTTAATAGCAGTGAAGGTGTGAGTGGCACCATCTTCTTT
Pea	ATGGTGAAGGGTGTGGCAGTCTTAGTAACAGTAACGAAGTCTCGGGTACTATTAACCTC
Arabidopsis	ATGGCGAAAGGAGTTGCAGTCTTTGAACAGCAGTGAAGGTGTGTTACGGGGACTATCTTTTTC
Oryza	ATGGTGAAGGGTGTGGCTGTTCTGCTAGCAGTGAGGGTGTCAAGGGCACCATCTTTTTC
	**** *
Malus	GTCCAGGAGGGAGATGGCCCAACTACTGTGACTGGAAGTGTCTCTGGCCTCAAGCCTGGA
Potentilla	ACCCAACAGCCGACATCCCCCAACTACTCTGACCCGAAACATTTCTCCCTCAACCCCTGGC
Populus	ACCCAAGAAGGAGATGGCCCAACTACTGTAAATTGGAACGTTTCTGGTGTAAAGCCAGGC
Pea	ACTCACCAGCCAAATCCTCCAACCCTCTAACTCCAACCTCTTCTCTCTTAAACCCCTGCC
Arabidopsis	ACCCAGGAAGGGGATGGTGTGACCACTGTGAGTGGAAACAGTTTCTGGCCTTAAAGCCTGGT
Oryza	TCCCAAGAGGGAGATGGTCCGACCTCTGTGACGGGAAGTGTCTCTGGGCTCAAGCCAGGG
	* *
Malus	CTTCATGGTTCATATGCTGCTCTTGGAGACACAACAAACGGTTGCATGTCAACTGGG
Potentilla	CTTCATGGTTCATATGCTCTTGGGGACACAACCAATGGTTGCATGTCAACTGGA
Populus	CTTCATGGCTTCCACGTCCATGCCCTTGGAGACACCACAATGGCTGCATGTCAACTGGG
Pea	CTCCACGGCTTCCATATCCATGCTTGGGAGACACCACAACGGTTGCATTTCAACTGGA
Arabidopsis	CTTCATGGTTCATATGCTCTTGGTGCACCACTAACCAGTTGCATGTCTACTGGT
Oryza	CTCCATGGATTCCATGTGCACGCGCTCGGTGACACCCTAATGGCTGCATGTCAACTGGA
	* *
Malus	CCACACTTCAATCCTGCTGGAAAAGAGCATGGTGCCCTGAAGATGAGCTTCGCCATGCT
Potentilla	CCACATTTCAATCCTGCTGGCAAAGAGCATGGGTCTCCTGAAGATGAGACTCGTCATGCT
Populus	CCGCATTTTAACTCTGTAGGCAAGGAGCATGGTGCCCTGAGGATGAGAACTGTCATGCT
Pea	CCACATTTCAATCCTAATGGGAAGGAACATGGTGCCCTGAGGATGAGACTAGACATGCT
Arabidopsis	CCACATTTCAACCCGATGGTAAACACACGGTGCCCTGAGGATGCTAATCGACATGCT
Oryza	CCACACTTCAATCCTACTGGGAAGGAACATGGGGCACCACAAGATGAGAACCAGCCATGCC
	* *
Malus	GGCGATCTTGGAAACATCAGTCTGGGGACGATGGAACGCAACCTTCACGATTGTTGAC
Potentilla	GGTGATCTTGGAAATATCAGTCTGGGGATGACGGAACTGCTTGTCTCACAATTGTTGAC
Populus	GGTGATCTGGGAAATGTCAGTCTGGTGTGATGATGACGCTGCTTTTCAAACTCATTGAC
Pea	GGTGATCTTGGAAATATCAATGTTGGTGTGATGGAACGTAAGCTTCACCATTAAGTAC
Arabidopsis	GGTGATCTAGGAAACATCAGTCTGGGAGATGATGGAACGTCACCTTCACAATCAGTAT
Oryza	GGTGATCTTGGAAATATAACAGCTGGAGCAGATGGTGTGCTAATGTCAATGTCTCTGAC
	* *
Malus	AAGCAGATTCCTCTCGCTGGACCACACTCTATCATTTGGTAGGGCGGTTGTTGTCCACGCA
Potentilla	AAACAGATTCCTCTCCTACTGGACCACACTCTATCATTTGGTAGGGCTGTTGTTGTCCATGCA
Populus	AAACAGATTCCTCTTACTGGACCACATTCATTTGGTTGGGCTGTTGTTGTTCCATGGA
Pea	AACCATATCCCTCTCCTACTGGAAACAACCTCCATCATAGGAAGGGCTGTTGTTGTCCATGCC
Arabidopsis	TGCCAGATTCCTCTTACTGGACCAACTCTATTGTTGGTAGGGCTGTTGTTGTCCATGCA
Oryza	AGCCAGATTCCTCTTACTGGAGCACAACCTCATCATTTGGCCGAGCTGTTGTTGTCCATGCT
	* *
Malus	GACCCTGATCACCTTCCCAACGCTGCACATGACCTTAGCAAATCCACACCAAATGCTGCT
Potentilla	GATCCTGATGACCTTGGCAAGGGTGGACATGAGCTTAGCAAATCCACTGGAATGCTGGT
Populus	GATCCTGATGATCTTGGCAAGGGAGGACATGAACCTCAGCAAACCACTGGAATGCTGGC
Pea	GATCCTGATGATCTTGGGAAAGGTGGTACGAGCTTAGCAAACCTACTGGAATGCTGGT
Arabidopsis	GACCCTGATGACCTCGGAAAGGGAGGCCATGAACCTCAGCTGCTACTGGAACCGCAGGC
Oryza	GATCCTGATGATCTTGGCAAGGGTGGACATGAGCTTAGCAAAGCCACTGGAATGCTGGG
	* *
Malus	GGCAGGGTGGCTTGGCGTATTATTGGTCTGCAAGGATGA
Potentilla	GGCAGGATAGCTTGTGGTATTATTGGCCTTCAAGGATGA
Populus	GGCAGAGTAGCATGCGGTATTATTGGTCTGCAAGGTTGA
Pea	GGCAGAGTAGCTTGTGGTATTATTGGTCTGCAAGGATGA
Arabidopsis	GGCCGTGTTGCTTGGCGCATATTGCTCTCCAGGGCTAA
Oryza	GGCCGAGTTGCTTGGGAATCATCGACTCCAGGGTTAG

Fig.3. Comparison of the deduced amino acid sequence of the *Potentilla* Cu/Zn SOD with sequences from other plant species. Regions of complete homology are indicated with asterisks.

Malus	MVKGVAVLGSSEGVKGTISFVQEGDGPPTTVTGSVSLKPLGLEGFHVHALGDTTNGCMSTG
Potentilla	MAKGVAVLSSSEGVAGTILFTQEGDGPPTTVGNISGLKPLGLEGFHVHALGDTTNGCMSTG
Arabidopsis	MAKGVAVLNSSEGVGTGTFEFTQEGDGPPTTVSGVSLKPLGLEGFHVHALGDTTNGCMSTG
Populus	MVKAVAVLNSSEGVSGTIFFTQEGDGPPTTVGNLSGLKPLGLEGFHVHALGDTTNGCMSTG
Cryza	MVKAVVVLGSSEIVKGTIFHVQEGDGPPTTVTGSVSLKPLGLEGFHIIHALGDTTNGCMSTG
Zea	MVKAVAVLGSSEFVQEGDGPPTTVTGSVSLKPLGLEGFHVHALGDTTNGCMSTG
Gossypium	MVKAVAVLGSSEGVSGTVFFSQQEGDGPPTTVGNLSGLKPLGLEGFHVHALGDTTNGCMSTG
Pisum	MVKAVAVLNSNEVSGTINFSQEGDGPPTTVTGTLAGLPLGLEGFHIIHALGDTTNGCISTG
Soybean	MVKAVAVLGSSEGVGTGTFEFTQEGDGPPTTVTGSVSLKPLGLEGFHVHALGDTTNGCLSTG
	* . *
Malus	PHFNPAGKEFGAPEDELRHAGDLGNITAGDDGTATFTIVDKQIPLAGPHSIIGRAVVVHIA
Potentilla	PHFNPAGKEFGSPEDETRHAGDLGNITVGGDGTACFTIVDKQIPLTGPHSIIGRAVVVHA
Arabidopsis	PHFNPDGKTFGAPEDANRHAGDLGNITVGGDGTATFTITDCQIPLTGPHSIVGRAVVVHA
Populus	PHFNPVGKFFGAPEDENRHAGDLGNITVGGDGTAAFTIIDFQIPLTGPHSIIGRAVVVHG
Cryza	PHYNPAGKEFGAPEDETRHAGDLGNVTAGDGVANIHVVDSQIPLTGPHSIIGRAVVVHA
Zea	PHYNPASKKEFGAPEDENRHAGDLGNVTAGDGVANINVTDSQIPLTGPHSIIGRAVVVHA
Gossypium	PHFNPAGKEFGAPEDENRHAGDLGNITVGGDGCASF SITDKQIPLTGPHSIIGRAVVVHA
Pisum	PHFNPNKKEFGAPEDETRHAGDLGNINVGGDGTVSEFTITDNEIPLTGTNSIIGRAVVVHA
Soybean	AHFNPNNNEFGAPEDENRHAGDLGNVNVGGDGTVSF SITDSQIPLTGPHSIIGRAVVVHA
	. * : * * . : * * : * * * * * . . . * * * . . : : * : * * * : * . : * * * * * * * .
Malus	DPDDLKGGGEELSSTGNAGGRVACGIIQLQG
Potentilla	DPDDLKGGGEELSSTGNAGGRVACGIIQLQG
Arabidopsis	DPDDLKGGGIELSLATGNAGGRVACGIIQLQG
Populus	DPDDLKGGGEELSKTIGNAGGRVACGIIQLQG
Cryza	DPDDLKGGGEELSKTIGNAGGRVACGIIQLQG
Zea	DPDDLKGGGFELSKSTGNAGGRVACGIIQLQG
Gossypium	DPDDLKGGGEELSSTGNAGGRVACGIIQLQG
Pisum	DPDDLKCCGEELSKTTCNACCRVACGIIQLQG
Soybean	DSDDLKGGGEELSKTIGNAGGRVACGIIQLQG
	* . * * * * * * * * * : * * * * * : * * * * * * * * *

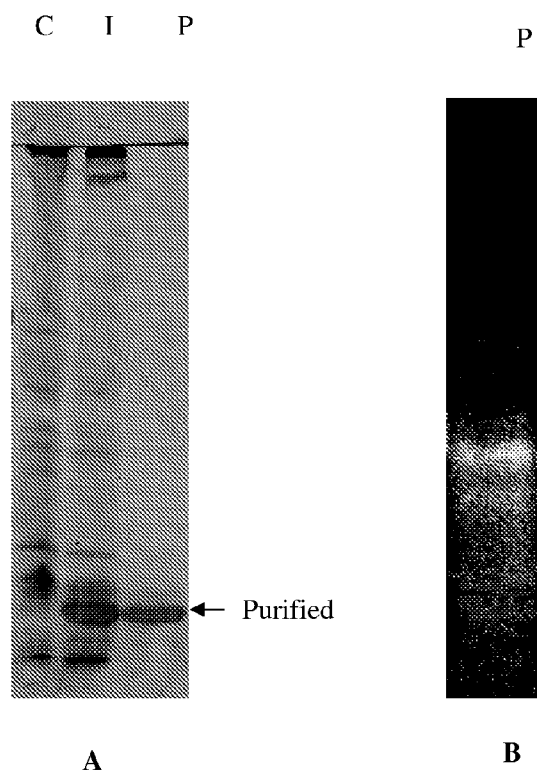


Fig.4. (A) Expression and purification of *Potentilla* SOD in *E. coli*. C, Control; I, Protein induced by IPTG; P, Purified SOD. The gel was stained by silver staining. (B) Activity staining of the gel to depict the activity of purified SOD. P, Purified SOD.

Fig-5: result of alignment of present sod gene with the sod gene of other plant species

Sequence 1: gi|311970|gi|311970I.batatas mRNA for superoxide dismutase
Length = 459 (1 .. 459)

Sequence 2: lcl|IHBT-potentilla
Length = 459 (1 .. 459)

Score = 348 bits (181), Expect = 8e-93
Identities = 323/394 (81%), Gaps = 0/394 (0%)
Strand=Plus/Plus

Query 56 TCTTCAGCCAAGAAGGAGATGGTCCAACCACAGTCACTGGAAACGTTTCGGGCCTCAAAC 115
Sbjct 56 TCTTTACCCAAGAGGGAGATGGCCCAACTACTGTGACCGGAAACATTCTGGCCTCAAGC 115
Query 116 CTGGTCTTCATGGCTTCCATGTCCATGCCCTAGGTSACACAACAAATGGATGCATGTCTA 175
Sbjct 116 CTGCCCTTCATGCTTCCATCTTCATGCTCTTCGGGACACAACCAATGCTTCCATGCTCAA 175
Query 176 CTGGACCACATTTCAATCCTGCTGGAAAGGAGCATGGAGCTCCTGGAGACGATAACCGCC 235
Sbjct 176 CTGGACCACATTTCAATCCTGCTGGCAAAGAGCATGGGTCTCCTGAAGATGAGACTCGTC 235
Query 236 ATGCCGGTGATCTTGGAAACATCACGGTTGGAGAAGATGGTACTGCTTCATTCACCATCA 295
Sbjct 236 ATGCTGGTGATCTTGGAAATATCACTGTTGGGGATGACGGAACTCCTGCTTCACAATTG 295
Query 296 CTGACAAGCAGATTCCGCTTACTGGAGCAAATCTGTTATTGGAACAGCTGTTGTGTTC 355
Sbjct 296 TTGACAAACAGATTCTCTCACTGGACCACACTTATCATTGGTAGGGCTGTTGTGTCC 355
Query 356 ATGGTGATCCCGATGATCTTGGTAAAGGTGGCCATGAGCTCAGCAAAAGCACTGGAAATG 415
Sbjct 356 ATGCAGATCCTCATGACCTTCGCAAGGGTGGACATGAGCTTAGCAAATCCACTCGAAATG 415
Query 416 CTGGCGGGAGGGTTGCCTGCGGTATCATTGGCCT 449
Sbjct 416 CTGGTGGCAGSATAGCTTGTGGTATTATTGGCCT 449

> gi|38228696|emb|AJ586519.1| Fagus sylvatica partial sod1 mRNA for superoxide dismutase
Length=710

Score = 379 bits (191), Expect = 5e 102
Identities = 392/459 (85%), Gaps = 0/459 (0%)
Strand=Plus/Plus

Query 1 ATGGCAAAGGGGCGTTGGCTGTACTTAGCTCCAGTGAGGGTGTGCTGGAACTATCCTCTTT 60
Sbjct 43 ATGGCCAAAGGGTGTGGCTGTTCTTAGCTCGAATGAGCGTGTGTTGTCGCACTATCTACTTT 102
Query 61 ACCCAAGAGGGAGATGGCCCAACTACTGTGACCGGAAACATTCTGCGCCTCAAGCCTGGG 120
Sbjct 103 GCCCAAGAGGAGATGGCCCAACTACAGTAACTGGAAATATTCTGCGCCTTAAACCTGGA 162
Query 121 CTTTCATGGTTTCCATGTTTCATGCTCTTGGGGACACAACCAATGGTTGCATGTCAACTGGA 180
Sbjct 163 CTCCATGGCTTCCACGTGCATGCTCTTGGGGACACAACAAATGGTTGCATGTCAACTGGA 222
Query 181 CCACATTTCAATCCTGCTGCCAAAGACCATGCTCTCCTCAACATCAGACTCCTCATGCT 240

Fig 5 Continued

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Sbjct 223  ||||| 282
Query 241  GGTGATCTTGAAATATCACTGTTGGGGATGACGGAACTGCTTGCTTCACAATTGTTGAC 300
          |||||  |||||  |||  |||||  |||||  ||  ||  |  |||||  |||||
Sbjct 283  GGTGATCTGGAAATGTCAATGTTGGTGTATGATGGCACAGTCAGTTTCACAATAATTGAC 342
Query 301  AAACAGATTCCTCTCACTGGACCACACTCTATCATTGGTAGGGCTGTTGTTGTCCATGCA 360
          |||||  ||  |||  ||  ||  ||  ||  ||  |||||  |||||  ||
Sbjct 343  AAACAGATTCACACTTTGTGGTCCAAATCCATTATCGGAAGGGCTGTTGTTGTCCATGGA 402
Query 361  GATCCTGATGACCTTGGCAAGGGTGGACATGAGCTTAGCAAATCCACTGGAAATGCTGGT 420
          |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||
Sbjct 403  GATCCAGATGATCTTGGCAAGGGGGACATGAAGTACTAGCAAGAGCACTGGAAATGCTGGT 462
Query 421  GGCAGGATAGCTTGTGGTATTATTGGCCTTCAAGGATGA 459
          |||  |  |||||  |||||  |||||  ||  |||||
Sbjct 463  CCCCCTATACCTTCTCTATCATTGCTCTCCAACGATCA 501

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> gi|4102858|gb|AF016892.1|AF016892 Populus tremuloides cytoplasmic superoxide dismutase
1 (SODcyt1)
mRNA, complete cds
Length=787

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Score = 333 bits (168), Expect = 3e-88
Identities = 342/400 (85%), Gaps = 0/400 (0%)
Strand=Plus/Plus

Query 56  TCTTTACCCAAGAGGGAGATGGCCCAACTACTGTGACCGGAAACATTTCTGGCCTCAAGC 115
          |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||
Sbjct 134  TCTTTACCCAAGAACACATGCCCAACTACTCTAATTGCAAACCTTTCTGCTCTTAACC 193

Query 116 CTGGGCTTCATGGTTTCCATGTTTCATGCTCTTGGGGACACAACCAATGTTGCATGTCAA 175
          |  ||  |||||  |||||  ||  |||||  |||||  ||  |||||  |||||  |||||
Sbjct 194  CAGGCTTCATGGCTTCCACGTCCATGCCCTTGGAGACACCACAAATGGCTGCATGTCAA 253

Query 176 CTGGACCACATTTCAATCCTGCTGGCAAAGAGCATGGGTCTCCTGAAGATGAGACTCGTC 235
          ||||  ||  |||||  |||||  |||||  |||||  ||  |||||  |||||  |||||
Sbjct 254  CTGGGCCGCAATTTAATCCTGTAGGCAAGGAGCATGGTGGCCCTGAGGATGAGAATCGTC 313

Query 236 ATGCTGCTGATCTTGGAAATATCACTGTTGGGGATGACGGAACTCCTTGCTTCACAATTG 295
          |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||
Sbjct 314  ATGCTGGTATCTGGAAATGTCACATGTTGGTGTATGATGGCACTGCTGCTTTCACAATCA 373

Query 296 TTGACAAACAGATTCCTCTCACTGGACCACACTCTATCATTGGTAGGGCTGTTGTTGTCC 355
          |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||
Sbjct 374  TTGACAAACAGATTCCTCTTACTGGACCACATTCCATTATGGTTGGGCTGTTGTTGTTC 433

Query 356 ATGCAGATCCTGATGACCTTGGCAAGGGTGGACATGAGCTTAGCAAATCCACTGGAAATG 415
          |||  |||||  |||||  |||||  |||||  |||||  ||  |||||  |||||  |||||
Sbjct 434  ATGGAGATCCTGATGATCTTGGCAAGGGGAGGACATGAACTCAGCAAACCCTGGTAATG 493

Query 416 CTGGTGGCAGCATAGCTTGTGGTATTATTGGCCTTCAAGG 455
          ||||  |||||  ||||  ||  |||||  |||||  ||  |||||
Sbjct 494  CTGGCGGCAGAGTAGCATGCGGTATTATTGGTCTGCAAGG 533

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> gi|50540928|gb|AY642137.1| Manihot esculenta copper/zinc superoxide dismutase mRNA,
complete
cds
Length=774

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Score = 333 bits (168), Expect = 3e-88
Identities = 354/416 (85%), Gaps = 0/416 (0%)
Strand=Plus/Plus

Query 31  AGTGAGGGTGTGCTGGAACATCCTCTTTACCCAAGAGGGAGATGGCCCAACTACTGTG 90
          |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||  |||||

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Fig 5 Continued

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Sbjct  44  ACTGAGGGTGTGCTGGGACAACTCTTCTTACCCCAAGAAGGAGATGGTCCAACCACCSTC 103
Query  91  ACCGGAAACATTTCTGGCCTCAAGCCTGGGCTTCATGGTTTCCATGTTTCATGCTCTTGGG 150
      || |||| | ||||| |||| | ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 104  ACTGCAACTCTTTCTCGCCTTAAGCCACCGCTTCATCGATTCCATGTTTCATCCCTTCCA 163
Query 151  GACACAACCAATGGTTGCATGTCAACTGGACCACATTTCAATCCTGCTGGCAAAGAGCAT 210
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 164  GACACAACAAATGGTTGCATGTCAACTGGGCCACATTTCAACCCTGGTGGCAAAGAGCAT 223
Query 211  GGGTCTCCTGAAGATGAGACTCGTCATGCTGGTGATCTTGAAAATATCACTGTTGGGGAT 270
      || | |||| | || | ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 224  GGTGCCCTGAGGACGACATTCGTCTGCTGGTGATCTGGGAAATGTCACCTGCTGGTGAT 283
Query 271  GACGSAACTGCTTGCTTACCAATTGTTGACAAACAGATTCTCTCACTGGACCACACTCT 330
      || | |||| | ||||| ||||| | ||||| |||| | || | || |
Sbjct 284  GATGGCACTGCTAGTTTACCAATCGTTGACAAGGATATTCTCTTTCTGGTCCGCATTCC 343
Query 331  ATCAITGGTAGGGCTGTTGTTGTCCATGCAGATCCTGATGACCTTGGCAAAGGGTGGACAT 390
      || | |||| | |||| | ||||| ||||| |||| | |||| | |||| |
Sbjct 344  ATTGTAGGAAGGGCAGTCGTTGTTTCATGCAGATCCTGATGATCTTGGAAAGGGGGACAT 403
Query 391  GAGCTTAGCAAAATCCACTGGAATGCTGGTGGCAGGATAGCTTGTGGTATTATTGG 446
      || ||||| ||||| ||||| ||||| |||| | |||| | |||| |
Sbjct 404  GAACCTAGCAAAACCCTGGAATGCTGGTGGCAGGGTAGCATGTGGTATTATTGG 459

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> gi|5723475|emb|AJ279694.1|BPE279694 Betula pendula partial mRNA for copper/zinc-
superoxide dismutase
(cu/Zn sod gene)
Length=355

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Score = 323 bits (163), Expect = 3e-85
Identities = 289/331 (87%), Gaps = 0/331 (0%)
Strand=Plus/Plus

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Query  49  ACTATCCTCTTTACCCAAGAGGGAGATGGCCCAACTACTGTGACCGGAAACATTTCTGGC 108
      ||||| ||||| ||||| | ||||| ||||| || || |||| | |||||
Sbjct  25  ACTATCCACTTTACCCAAGAAGCTGATGGCCCAACTACAGTAACTGGAAATATTTCTGGC  84
Query 109  CTCAGCCTGGGCTTCATGGTTCCATGTTTCATGCTCTTGGGGACACAACCAATGGTTGC 168
      || ||||| |||| | ||||| |||| | ||||| ||||| ||||| |||||
Sbjct  85  CTTAACCCTCCCTCCATCCGTTCCATCTCCATCCACTTCGGCACACAACAAATCCTTCC 144
Query 169  ATGTCAACTGGACCACATTTCAATCCTGCTGGCAAAGAGCATGGGTCTCCTGAAGATGAG 228
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 145  ATGTCAACTGGGCCACATTTCAATCCTGCTGGCAAAGAGCATGGTCTCCTGAGGATGAG 204
Query 229  ACTCGTCATGCTGGTGATCTTGGAAAATATCACTGTTGGGGATGACGGAACTGCTTGCTTC 288
      | ||||| ||||| ||||| |||| | |||| | |||| | |||| | |||| | ||||
Sbjct 205  AATCGTCATGCCGGTGATCTGGGAAATGTCACCGTTGGTGATGATGGTACTGCCAGTTTC 264
Query 289  ACAATTGTTGACAACAGATTCTCTCACTGGACCACACTCTATCATTGGTAGGGCTGTT 348
      |||| | ||||| ||||| || | ||||| |||| | |||| | |||||
Sbjct 265  ACAATAGTTGACAAGCAGATTCCACTTTCTGCACCACATTCTATTATTGGAAGGGCTGTT 324
Query 349  GTTGTCATGCAGATCCTGATGACCTTGGCA 379
      ||||| | |||| | |||| | |||||
Sbjct 325  GTTGTCACGGGGATCCAGATGATCTTGGCA 355

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> gi|13274149|emb|AJ278669.1|PTR278669 Populus tremula x Populus tremuloides mRNA for
putative cytosolic
CuZn superoxide dismutase (cyt SOE1 gene)
Length=851

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Score = 305 bits (154), Expect = 6e-80
Identities = 331/390 (84%), Gaps = 0/390 (0%)
Strand=Plus/Plus

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Fig 5 Continued


Query 57 CTTTACCCAAGAGGAGATGGCCCAACTACTGTGACCGGAAACATTTCTGGCCTCAAGCC 116
Sbjct 135 CTTTACCCAAGAAGGAGATGGTCCAACTACTGTAAGTGGAAAGCCTCTGTGGTCTTAAGCC 194
Query 117 TGGGCTTCATGGTTTCCATGTTTCATGTCTTGGGGACACAACCAATGGTTGCATGTCAAC 176
Sbjct 195 AGGCCTTCATGGCTTCCATGTTTCATGCCCTTGGAGACACCACAAATGGCTCCATGTCAAC 254
Query 177 TGGACACATTTCAATCCTGCTGGCABAGAGCATGGGTCTCCTGAAGATGAGACTCGTCA 236
Sbjct 255 TGGCCCGCATTTTTAATCCTGTAGGCABAGAGCATGGTGGCCCTGAGGATGAGAATCGTCA 314
Query 237 TGCTGGTGTATCTTGGAAATATCACTGTTGGGGATGACGGAACTGCTTGCTTACAAATGT 296
Sbjct 315 TGCTGGTGTATTTGGGAAATGTCACTCTTGGTGTATGATGGCACCCTACTGTCTCAATCAT 374
Query 297 TGACAAACAGATTCTCTCACTGGACCACACTCTATCATTGGTAGGGCTGTTGTTGTCCA 356
Sbjct 375 TGACAACAGATTCTCTCACTGGACCAGAAATCCATCGTGGAAAGGGCTGTTGTTGTCCA 434
Query 357 TGCAGATCCTGATGACCTTGGCAAGGGTGGACATGAGCTTAGCAAAATCCACTGGAAATGC 416
Sbjct 435 TGCAGATCCTGATGATCTTGGCAAGGGAGGACATGAACTTACCAAAAAGCACTGGTAATCC 494
Query 417 TCCTCCACACATACCTTCTCCTATTATTCC 446
Sbjct 495 TGGTGGCAGAGTAGCATGTGGTATTATTGG 524

[] >gi|53748478|emb|A7844003.1| Plantago major mRNA for copper-zinc superoxide dismutase (csd1 gene) Length=779

Score = 305 bits (154), Expect = 6e-80
Identities = 373/446 (83%), Gaps = 0/446 (0%)
Strand=Plus/Plus

Query 7 AAGGGCGTTGCTGTACTTAGCTCCAGTGAGGGTGTGCTGGAACTATCCTCTTTACCCAA 66
Sbjct 70 AAGGGTGTTCAGTGTCTTAGCAGCAGTGAGGGTGTAGTGGCACCCTCTTTTCCCAA 129
Query 67 GAGGGAGATGGCCCAACTACTGTGACCGGAAACATTTCTGGCCCAAGCCTGGGCTTCAT 126
Sbjct 130 CAAGGAGAAGGACCCACTGTAAGTGGAAACCTTTCTGGCCCAAGCCTGGACTTCAC 189
Query 127 GGTTCATGTTTCATGCTCTTGGGGACACAACCAATGGTTGCATGTCAACTGGACCACAT 186
Sbjct 190 GGCTTCCATGTTTCATGCTCTTGGTGTGACTACCAACGGTTCATGTCAACAGGACCACAT 249
Query 187 TTCAATCCTTGGTGGCAAGAGCATGGGTCTCCTGAAAGATGAGACTCGTTCATGCTGGTGTAT 246
Sbjct 250 TTCAATCCGCTGCAAAAAGAGCATGCTGCTCCTCATGATGAGCTTCGCCATGCTGGTGTAC 309
Query 247 CTTGGAATATCACTGTTGGGGATGACGGAACTGCTTGTTCACAATTGTTGACAAACAG 306
Sbjct 310 CTTGGTAATGTCACAGTGGGAGATGATGGAAGTTCACCATTGTTGACAAGCTG 369
Query 307 ATTCCTCTCACTGGACCACACTCTATCATTGGTAGGGCTGTGTTGTCATGCAGATCCT 366
Sbjct 370 ATTCGGCTGACTGGACCACATTCATCATTGGAAGGGCTGTTGTTGTCATGCTGACCCC 429
Query 367 GATGACCTTGGCAAGGGTGGACATGAGCTTAGCAAAATCCACTGAAATGCTGGTGGCAGG 426
Sbjct 430 GATGATTTGGGAAGGGTGGACATGAACTCAGCAAAAATACCGAAATGCTGGTGGGAAGA 489
Query 427 ATAGCTTGTGTTATTATTGGCCCTTCA 452
Sbjct 490 GTTGCTTGTGTATCATTTGGTCTTCA 515

Fig 5 Continued

 [gi|5726591|gb|AF170297.1|AF170297](#) Manihot esculenta copper/zinc-superoxide dismutase mRNA, complete cds
Length=801

Score = 295 bits (149), Expect = 6e-77
Identities = 338/401 (84%), Gaps = 0/401 (0%)
Strand=Plus/Plus

```

Query 46  GGAACATATCCTCTTTACCCAAGAGGGAGATGGCCCAACTACTGTGACCGGAAACATTCT 105
          ||||| ||| ||||| ||||| ||||| ||| || ||||| || ||||| |||||
Sbjct 100  GGAACAATCTTCTTTACCCAAGAAGGAGATGGTCCTACCACTGTAACGGAAACATTTC 159

Query 106  GGCCTCAACCCCTGGCCTTCATGCTTTCCATGCTTCATGCTCTTCGGCACACAACCAATGCT 165
          ||||| ||||| ||||| ||||| ||||| || ||||| ||||| ||||| || |||
Sbjct 160  GGCCTTAAGCCAGGGCTTCATGGGTTCCACGTCATGCCCTTGGAGACACAACAACGGT 219

Query 166  TGCATGTCAACTGGACCACATTTCAATCCTGCTGGCAAAGAGCATGGGTCTCCTGAAGAT 225
          ||||| ||||| ||||| || || || || ||||| ||||| || ||||| |||
Sbjct 220  TGCATGTCAACTGGGCCACACTTTAACCCCTTCTGGCAAAGATCATGGTGCCTCTGAGGAT 279


Query 226  GAGACTCGTCATGCTGGTGATCTTGCAAATATCACTGTTGGGCATGACGGAACCTGCTTCC 285
          ||||| ||||| ||||| ||||| ||||| || ||||| || ||||| || ||||| |
Sbjct 280  GAGATTCGTCATGCTGGTGATCTGGGAAATGTCACCTGCTGGTGATGATGGCACTGCTAGT 339

Query 286  TTCACAATTGTTGACAAACAGATTCTCTCACTGGACCACACTCTATCATTGGTAGGGCT 345
          ||||| ||||| || ||||| || ||||| || || || || ||||| || |||||
Sbjct 340  TTCACAATTATGACAAGCATATTCCTCTTTCTGGTCAAAATTCATCATAGGAAGGGCA 399

Query 346  GTTGTGTCCATGCAGATCCTGATGACCTTGGCAAAGGGTGGACATGAGCTTAGCAAATCC 405
          ||||| ||||| ||||| ||||| ||||| || ||||| || || || || ||
Sbjct 400  GTTGTGTTCATGCAGATCCTGATGATCTTGGCAGGGGAGGACATGAACTCAGTAAAACC 459

Query 406  ACTGGAATGCTGGTGGCAGGATAGCTTGTGGTATTATTGG 446
          || ||||| ||||| ||||| || ||||| |||||
Sbjct 460  ACCGGAATGCTGGTGGCAGAGTAGCATGCGGTATTATTGG 500

```

 [gi|56549630|gb|AY833718.1|](#) Codonopsis lanceolata CuZn superoxide dismutase (SODCc) mRNA, complete cds
Length=799

Score = 289 bits (146), Expect = 4e-75
Identities = 335/398 (84%), Gaps = 0/398 (0%)
Strand=Plus/Plus

```

Query 58  TTTACCCAAGAGGGAGATGGCCCAACTACTGTGACCGGAAACATTCTGGCCTCAAGCCT 117
          ||||| ||||| ||||| ||||| || || || || ||||| || ||||| || |||
Sbjct 212  TTTACCCAAGAGGGAGATGGCCCAACTAAAGTACTGGAAGCCTTCTGGCCTTCAACCT 271

Query 118  GCGCTTCATGCTTTCCATCTTCATGCTCTTCCGCACACAACCAATGCTTGCATGTCAACT 177
          || | ||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 272  GGACCTCACGGTTTCCATCTTCATGCCCTTGGTGACACAACCAATGGTTGCATGTCAACT 331

Query 178  GGACCACATTTCAATCCTGCTGGCAAAGAGCATGGGTCTCCTGAAGATGAGACTCGTCAT 237
          || || |||| | ||||| ||||| ||||| ||||| || || || || |||||
Sbjct 332  GGTCTCATTATAATCCTGCTGGAAAAGAACATGGTGCTCCAGAGGACGAGATTCTGTCAT 391

Query 238  GCTGGTGATCTTGGAAATATCACTGTTGGGGATGACGGAACCTGCTTGCATCACAATTGTT 297
          ||||| || || || || || || || || ||||| ||||| ||||| || |||
Sbjct 392  GCTGGTGACCTCGGAATGTTACAGTAGGCGAAGACGGTACTGCAAATTCACCATCGTT 451

Query 298  GACAAACAGATTCTCTCACTGGACCACACTCTATCATTGGTAGGGCTGTTGTTGTCCAT 357
          ||||| ||||| || ||||| || ||||| ||||| ||||| ||||| |||||
Sbjct 452  GACAACCAGATTCACATCTGGACCTCATTCTATCATTTGGAAGGGCTGTAGTTGTCCAT 511

Query 358  GCAGATCCTCATGACCTTGGCAAAGGCTGGACATGAGCTTAGCAAATCCACTGGAAATGCT 417

```


Fig 5 Continued

```

Query 177 TGGACCACATTTCAATCCTGCTGGCAAAGAGCATGGGTCTCCTGAAGATGAGACTCGTCA 236
      ||| || ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 274 TGGCCCGCATTTTAATCCTGTAGGCAAAGAGCATGGTGGCCCTGAGGATGAGAATCGTCA 333

Query 237 TGCTGGTGTATCTTGGAAATACACTGTTGGGGATGACGGAAGTCTTGCTTCACAATTGT 296
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 334 TGCTGGTGTATTGGGAAATGTCAGTGTGGTGTATGATGGCACCCTACTGTCICAATCAT 393


Query 297 TGACAAACAGATTCCCTCCTCACTGGACCACACTCTATCATTGGTAGGGCTGTTGTTGTCCA 356
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 394 TGACAACCAGATTCCCTTACTGGACCAAATTCATTGTTGGAAGGGCAGTTGTTGTTCA 453

Query 357 TGCACATCCTGATCACCTTGCCAAAGGCTGCACATGACCTTAGCAAATCCACTGCAAATGC 416
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 454 TGCAGATCCTGATGATCTTGGCAAGGGAGGACATGAACCTAGCAAAGCAGTGGTAATGC 513

Query 417 TGGTGGCAGGATAGCTTGTGGTATTATTGG 446
      ||||| ||||| ||||| ||||| |||||
Sbjct 514 TGGTGGCAGAGTAGCATGTGGTATTATTGG 543

```

```

> gi|27088051gb|AF037359.1|AF037359 Paulownia kawakamii superoxide dismutase (SOD5) mRNA,
complete
cds
Length=794

```

```

Score = 276 bits (139), Expect = 6e-71
Identities = 352/423 (83%), Gaps = 0/423 (0%)
Strand=Plus/Plus

```

```

Query 30 CAGTGAGGGTGTGCTGGAACTATCCTCTTTACCCAAGAGGGAGATGGCCCAACTACTGT 89
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 117 CAGTGAGGGTGTAGTGGCACCATCTACTTCACCCAGGAAGGAGATGGTCCAACAACACTGT 176

Query 90 CACCCCAAACATTTCTGCCCTCAAGCCTCCGCTTCATGCTTTCCATGTCATCCTCTTCC 149
      || ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 177 TACTGGAAACGTTTCTGGCCTTAAGCCTGGACCCCATGGCTTTCATGTCATGCCCTTGG 236

Query 150 GGACACAACCAATGGTTGCATGTCAACTGGACCACATTTCAATCCTGCTGGCAAAGAGCA 209
      ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 237 TGACACCACCAATGGTTGTTGTCAACTGGACCTCACTTCAATCCTGCTGGCAAAGAGCA 296

Query 210 TGGGTCTCCTGAAGATGAGACTCGTCATGCTGGTGTATCTTGGAAATACACTGTTGGGGA 269
      ||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 297 TGGAGCTCCTGATGATGAGGTTCCGCCATGCTGGTACCTTGGGAATGTCACAGTTGGAGA 356

Query 270 TGACGGAACTGCTTGCTTCACAATTGTTGACAAACAGATTCCCTCCTCACTGGACCACACTC 329
      || || ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 357 AGATGGCACTGCTGCTTTCATATTGTTGACAAGCAGATACCACTTACAGGACCACATTC 416

Query 330 TATCATTGGTAGGGCTGTTGTTGTCATGCAGATCCTGATGACCTTGGCAAAGGGTGGACA 389
      || ||||| || ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 417 CATAATTGGAAGAGCTGTAGTTGTTTCATGCTGATCCTGATGATCTTGGAAAGGGTGGACA 476


Query 390 TGAGCTTAGCAAATCCACTGGAATGCTGGTGGCAGGATAGCTTGTGGTATTATTGGCCT 449
      ||| || ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 477 TGAAGTGGCAAACCACTGGAATACTGGTGGAAAGAGTTGCTTGTGGTATCAATGGCCT 536

Query 450 TCA 452

      |||
Sbjct 537 TCA 539

```

```

> gi|398407781emb|AJ428575.2|OEU428575 Olea europaea Cu/Zn super-oxide dismutase (ole e 5
allergen)
Length=714

```

Fig 5 Continued

Score = 272 bits (137), Expect = 9e-70
Identities = 331/393 (84%), Gaps = 2/393 (0%)
Strand=Plus/Plus

Query 61 ACCCAAGAGGGAGATGGCCCAACTACTGTGACCGAAACATTTCTGGCCTCAAGCCTGGG 120
Sbjct 61 ACCCAAGAAGGAGATGGTCCAACTACTGTTACTGGAAACCTTTCTGGCCTTAAGCCTGGA 120
Query 121 CTTTCATGCTTTCCATGTTTCATGCTCTTGGGGACACAACCAATGGTTGCATGTCAACTGGA 180
Sbjct 121 CTTTCATGCTTTTCATGTTCCACGCCCTTGGTGACACCACCAATGGCTGTATGTCAACTGGA 180
Query 181 CCACATTTCAATCCTGCTGGCAAAGAGCATGGGTCTCCTGAAGATGAGACTCGTCATGCT 240
Sbjct 181 CCTCATTTCATCCTGTTGGGAAAGAGCATGGTGCACCTGGAGATGAGAACCCTCATGCT 240
Query 241 GGTGATCTTGGAAATATCACTGTTGGGGATGACGGAACCTGCTTCTCT-TCACAATTCTTGA 299
Sbjct 241 GGTGATCTTGGTAATATCACAGTTGGCGAAGATGGCACC GC -TGCTATCAACATTGTTGA 299
Query 300 CAAACAGATTCTCTCACTEGACCACACTCTATCATTGGTAGGGCTGTTGTTGTCCATGC 359
Sbjct 300 CAAGCAGATACCTCTTACAGGACCACATTCCATAATTGGAAGAGCAGTAGTTGTCCATTC 359
Query 360 AGATCCCTGATGACCTTGGCAAGGGTGGACATGAGCTTAGCAAATCCACTGGAATGCTGG 419
Sbjct 360 AGATCCCTGATGATCTTGGAAAGGGTGGTCATGAACTGAGCAAGAGCACTGGAATGCTGG 419
Query 420 TGGCAGGATAGCTTGTGGTATTATTGGCCTTCA 452
Sbjct 420 TGGAAGAGTTGCTTGTGGTATCATTGGCCTTCA 452

Sequence alignment header for Solanum tuberosum Cu/Zn-superoxide dismutase mRNA, partial cds. Includes a small square icon and a large empty rectangular box.

Score = 268 bits (135), Expect = 1e-68
Identities = 330/395 (83%), Gaps = 0/395 (0%)
Strand=Plus/Plus

Query 52 ATCCTCTTTACCCAAGAGGGAGATGGCCCAACTACTGTGACCGAAACATTTCTGGCCTC 111
Sbjct 30 ATCCTCTTCACTCAAGATGGAGATGCTCCAACCACAGTTAATGGAATATTTCTGGCCTA 89
Query 112 AAGCCTGGGCTTCATGCTTTCCATGTTTCATGCTCTTGGGGACACAACCAATGGTTGCATG 171
Sbjct 90 AAACCTGGACTTCATGGCTTCCATGTCCATGCCCTTGGTGATACCACAAATGGCTGCATG 149
Query 172 TCAACTGGACCACATTTCAATCCTGCTGGCAAAGAGCATGGGTCTCCTGAAGATGAGACT 231
Sbjct 150 TCAACAGGACCACATTACAATCCTGCTGGTAAGGAGCATGGTCTCCTGAAGATGAGGTG 209
Query 232 CGTCATGCTGGTGATCTTGGAAATATCACTGTTGGGGATGACGGAACCTGCTTGCCTCACA 291
Sbjct 210 CGTCATGCTGGTGATCTTGGTAACATCACAGTTGGAGAAGATGGTACTGCATCTTTACT 269
Query 292 ATTGTTGACAAACAGATTCTCTCACTGGACCACACTCTATCATTGGTAGGGCTGTTGTT 351
Sbjct 270 ATTACCACAAGCAGATTCTCTCACTGGTTCACAATCCATCATTGGAAGAGCTGTTGTT 329
Query 352 GTCCATGCAGATCCTGATGACCTTGGCAAGGCTGGACATGAGCTTAGCAAATCCACTGGA 411

Fig 6: Detail of SEQ ID No. 1

Organization Applicant

Street : Rafi marg,

City : New Delhi

State : Delhi

Country : India

PostalCode : -110001

PhoneNumber :

FaxNumber :

EmailAddress :

<110> OrganizationName : COUNCIL OF SCIENTIFIC & INDUSTRIAL RESEARCH

Application Project

<120> Title : SUPEROXIDE DISMUTASE (SOD) GENE AND A METHOD OF
IDENTIFYING AND
CLONING THEREOF

<130> AppFileReference : 0038NF2006

<140> CurrentAppNumber :

<141> CurrentFilingDate : 2006-03-31

Sequence

<213> OrganismName : Potentilla atrosanguinea

Fig 6 Continued

<400> PreSequenceString :

MAKGVAVLSS SEGVAGTILF TQEGDGPTTV TGNISGLKPG LHGFHVHALG

DTTNGCMSTG 60

PHFNPAGKEH GSPEDETRHA GDLGNITVGD DGTACFTIVD KQIPLTGPHS

IIGRAVVVHA 120

DPDDLKGGH ELSKSTGNAG GRIACGIIGL QG 152

<212> Type : PRT

<211> Length : 152

SequenceName : Polypeptide sequence of SOD gene SEQ ID NO. 1

SequenceDescription :

Fig 7: Detail of SEQ ID No. 2

Organization Applicant

Street : Rafi marg,

City : New Delhi

State : Delhi

Country : India

PostalCode : -110001

PhoneNumber :

FaxNumber :

EmailAddress :

<110> OrganizationName : COUNCIL OF SCIENTIFIC & INDUSTRIAL RESEARCH

Application Project

<120> Title : SUPEROXIDE DISMUTASE (SOD) GENE AND A METHOD OF IDENTIFYING AND

CLONING THEREOF

<130> AppFileReference : 0038NF2006

<140> CurrentAppNumber :

<141> CurrentFilingDate : 2006-03-31

Sequence

<213> OrganismName : Potentilla atrosanguinea

<400> PreSequenceString :

acgggggggg gactgaaata aatagagagg gtcatagtc cattgcatt taggtatctg 60

Fig 7 Continued

attccattca caaacctcca acteccacct ctctctctat ttctcttcat ctccatcacc 120
ttaggggtgca ctgagatcac ttgaaacat ggcaaagggc gttgctgtac ttagctccag 180
tgagggtgtt gctggaacta tctctttac ccaagaggga gatggcccaa ctactgtgac 240
cggaacatt tctggcctca agcctgggct tcatggtttc catgttcatg ctcttgggga 300
cacaaccaat gtttgcattg caactggacc acatttcaat cctgctggca aagagcatgg 360
gtctctgaa gatgagactc gtcattgctgg tgatcttggga aatcactg ttggggatga 420
cggaactgct tgcttcaaa ttgttgaaa acagattcct ctactggac cacactctat 480
cattggtagg gctgtgttg tccatgcaga tctgatgac ctggcaagg gtggacatga 540
gcttagcaaa tccactggaa atgctgggtg caggatagct tgtggtatta ttggcctca 600
aggatgaact ggaccagga gcgaacaca ggcatctgt tgaattaaaa ctgagatat 660
tagcgaactc ttcggaattg agtattgaaa caaggaatac attgtcatt accaatacgt 720
ttgcttaga cctgtattct gtatctcaat agtttctgt gtggtgttt gacagtatt 780
tgtctcagg ctattcaaa gggataaca cagtaacttt ctgcttga caaaaaaaaa 840
aaaaaaaaa aaaaaa 856

<212> Type : DNA

<211> Length : 856

SequenceName : Full length cDNA SOD gene of SEQ ID No. 2

SequenceDescription :

Fig 8: detail of SEQ ID No. 3

Organization Applicant

Street : Rafi marg,

City : New Delhi

State : Delhi

Country : India

PostalCode : -110001

PhoneNumber :

FaxNumber :

EmailAddress :

<110> OrganizationName : COUNCIL OF SCIENTIFIC & INDUSTRIAL RESEARCH

Application Project

<120> Title : SUPEROXIDE DISMUTASE (SOD) GENE AND A METHOD OF IDENTIFYING AND CLONING THEREOF

<130> AppFileReference : 0038NF2006

<140> CurrentAppNumber :

<141> CurrentFilingDate : 2006-03-31

Sequence

<213> OrganismName : Potentilla atrosanguinea

<400> PreSequenceString :

atggcaaagg gcgttgctgt acttagctcc agtgagggtg ttgctggaac tatcctcttt 60

Fig 8 Continued

acccaagagg gagatggccc aactactgtg accggaacaa ttctggcct caagcctggg 120
cttcatggtt tccatgttca tgctctggg gacacaacca atggttgcac gtcaactgga 180
ccacatttca atctgtctgg caaagagcat gggctcctg aagatgagac tcgtcatgct 240
ggtgatcttg gaaatatcac tgttgggat gacggaactg cttgcttcac aattgttgac 300
aaacagattc ctctactgg accacactct atcattggta gggctgtgtg tgcctatgca 360
gatcctgatg acctggcaa gggggacat gagcttagca aatccactgg aaatgctggt 420
ggcaggatag cttgtggtat tattggcctt caaggatga 459

<212> Type : DNA

<211> Length : 459

SequenceName : Coding sequence of potentialla SOD gene of SEQ ID NO. 3

SequenceDescription :

Fig 9: Detail of SEQ ID No. 4

Organization Applicant

Street : Rafi marg,

City : New Delhi

State : Delhi

Country : India

PostalCode : -110001

PhoneNumber :

FaxNumber :

EmailAddress :

<110> OrganizationName : COUNCIL OF SCIENTIFIC & INDUSTRIAL RESEARCH

Application Project

<120> Title : SUPEROXIDE DISMUTASE (SOD) GENE AND A METHOD OF IDENTIFYING AND

CLONING THEREOF

<130> AppFileReference : 0038NF2006

<140> CurrentAppNumber :

<141> CurrentFilingDate : 2006-03-31

Sequence

<213> OrganismName : Potentilla atrosanguinea

<400> PreSequenceString :

caagagggag atggccaac tactgtgacc ggaaacattt ctggcctcaa gcctgggctt 60

Fig 9 Continued

catggttcc atgttcatgc tcttggggac acaaccaatg gttgcatgtc aactggacca 120
catttcaatc ctgctggcaa agagcatggg tctctgaag atgagactcg tcatgctggt 180
gatcttggaa atatcactgt tggggatgac ggaactgctt gcttcacaat tgttgacaaa 240
cagattcctc tcaactggacc acactctatc attggtaggg ctgttggtt ccatgcagat 300
cctgatgacc ttggcaaggg tggacatgag cttagcaaat ccaactggaaa tgctggtggc 360
aggat 365

<212> Type : DNA

<211> Length : 365

SequenceName : Positive cDNA clone of SEQ ID No. 4

SequenceDescription :

Fig 10: Detail of SEQ ID No. 4

Organization Applicant

Street : Rafi marg,

City : New Delhi

State : Delhi

Country : India

PostalCode : -110001

PhoneNumber :

FaxNumber :

EmailAddress :

<110> OrganizationName : COUNCIL OF SCIENTIFIC & INDUSTRIAL RESEARCH

Application Project

<120> Title : SUPEROXIDE DISMUTASE (SOD) GENE AND A METHOD OF IDENTIFYING AND

CLONING THEREOF

<130> AppFileReference : 0038NF2006

<140> CurrentAppNumber :

<141> CurrentFilingDate : 2006-03-31

Sequence

<213> OrganismName : Potentilla atrosanguinea

<400> PreSequenceString :

atggcaaagg gcggtgctgt actt

Fig 10 Continued

<212> Type : DNA

<211> Length : 24

SequenceName : Primer Sequence SEQ ID No. 5(a) Forward Primer

SequenceDescription :

Sequence

<213> OrganismName : *Potentilla atrosanguinea*

<400> PreSequenceString :

tcatccttga aggccaataa tacca

25

<212> Type : DNA

<211> Length : 25

SequenceName : Primer sequence SEQ ID No. 5(b) : Reverse primer

SequenceDescription :

Fig 11: SEQ ID No. 6

Organization Applicant

Street : Rafi marg,

City : New Delhi

State : Delhi

Country : India

PostalCode : -110001

PhoneNumber :

FaxNumber :

EmailAddress :

<110> OrganizationName : COUNCIL OF SCIENTIFIC & INDUSTRIAL RESEARCH

Application Project

<120> Title : SUPEROXIDE DISMUTASE (SOD) GENE AND A METHOD OF IDENTIFYING AND

CLONING THEREOF

<130> AppFileReference : 0038NF2006

<140> CurrentAppNumber :

<141> CurrentFilingDate : 2006-03-31

Sequence

<213> OrganismName : Potentilla atrosanguinea

<400> PreSequenceString :

ccagtggatt tgctaagetc atgtcca

27

<212> Type : DNA

Fig 11 Continued

<211> Length : 27

SequenceName : Primer Sequence of SEQ ID No. 6(a) GSP1:Forward primer

SequenceDescription :

Sequence

<213> OrganismName : *Potentilla atrosanguinea*

<400> PreSequenceString :

gtcatcaggg tctgcatgga caacaac 27

<212> Type : DNA

<211> Length : 27

SequenceName : Primer sequence of SEQ ID No. 6(b) NES1: Reverse Primer

SequenceDescription :

Sequence

<213> OrganismName : *Potentilla atrosanguinea*

<400> PreSequenceString :

atggttgcac gtcaactgga ccacatt 27

<212> Type : DNA

<211> Length : 27

SequenceName : Primer Sequence of SEQ ID No. 6(c) GSP2: Forward Primer

SequenceDescription :

Sequence

<213> OrganismName : *Potentilla atrosanguinea*

<400> PreSequenceString :

ttgcatgtca actgaccac atttcaa 27

<212> Type : DNA

Fig 11 Continued

<211> Length : 27

SequenceName : Primer sequence of SEQ ID No. 6(d) NES2: Reverse Primer

SequenceDescription :

Sequence

<213> OrganismName : *Potentilla atrosanguinea*

<400> PreSequenceString :

aagcagtggg atcaacgcag agtacgcggg 30

<212> Type : DNA

<211> Length : 30

SequenceName : Primer Sequence of SEQ ID No. 6(e): SMART II A Oligonucleotide

SequenceDescription :

Sequence

<213> OrganismName : *Potentilla atrosanguinea*

<400> PreSequenceString :

aagcagtggg atcaacgcag agtactnn 28

<212> Type : DNA

<211> Length : 28

SequenceName : Primer sequence of SEQ ID No. 6(f): 3'- RACE CDS Primer A (3'-
CDS):

SequenceDescription :

1

**SUPEROXIDE DISMUTASE (SOD) GENE AND
A METHOD OF IDENTIFYING AND
CLONING THEREOF**

CROSS-REFERENCE TO RELATED
APPLICATIONS

This application is a divisional of U.S. patent application Ser. No. 11/499,505, filed Aug. 4, 2006, which claims benefit of Indian application 0928/DEL/2006, filed Mar. 31, 2006. The contents of each of these applications are incorporated herein by reference in their entirety.

FIELD OF INVENTION

The present invention relates to a Superoxide dismutase (SOD). Superoxide dismutase (SOD) cDNA of SEQ ID No. 2 obtained from *Potentilla atrosanguinea* containing coding gene sequence of SEQ ID No. 3 which codes for a polypeptide of SEQ ID No. 1 having Superoxide dismutase enzyme activity.

Further, it also relates to a set of primers useful for the amplification of Superoxide dismutase (SOD) gene coding cDNA of SEQ ID No. 3, wherein

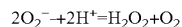
Forward primer (SEQ ID NO: 5)
5'-ATGGCAAAGGGCGTTGCTGTA CTT-3'
and;

Reverse primer (SEQ ID NO: 6)
5'-TCATCCTTGAAGGCCAATAATACCA-3'

More particularly, it relates to a method of identifying and cloning of Superoxide dismutase (SOD) gene of SEQ ID NO 3, which on expression gives a Superoxide dismutase enzyme (EC 1.15.1.1) with the characteristics disclose in U.S. Pat. No. 6,485,950.

BACKGROUND AND PRIOR ART REFERENCES
TO THE INVENTION

SOD is a ubiquitous enzyme present in plants, animals and microbes, which protects them against oxidative damage caused by superoxide radical (hereinafter, referred to O₂⁻). The enzyme dismutates superoxide radical into hydrogen peroxide and oxygen as per the following redox reaction:



Thus, SOD has implications in all those reactions, wherein O₂⁻ is produced in the amount leading to cellular injury.

2

According to the U.S. Pat. No. 6,485,950, we have extracted an autoclavable superoxide dismutase from *Potentilla* that could be autoclaved and shows activity at sub-zero temperature. Due to prevalence of *Potentilla* at difficult to access location of high altitude, and industrial implications of SOD as mentioned in our U.S. Pat. No. 6,485,950, it was essential to develop a system for the production of SOD of *Potentilla* in *E. coli* so as to obtain the SOD when desired.

Below is given state of the art knowledge in relation to isolation of SOD genes from various sources and their expression in *E. coli*, to produce SOD in recoverable quantities.

Reference may be made to document (1) by Wang, Z., He, Z., Shen, Q., Gu, Y., Li, S, and Yuan, Q. (J. of Chromatography B, 2005. 826: 114-121) wherein Cu/Zn SOD gene from *Cordyceps militaris* was overexpressed in *E. coli*.

Yet another reference may be made to document (2) by Liu, W., Zhu, R. H., Li, G. P., and Wang, D. C. (Protein Expr. Purif. 2002. 25: 379-388) wherein production of high yield of recombinant duck Cu/Zn SOD was achieved in *E. coli*.

Reference may be made to yet another document (3) by Pan, S. M., Hwang, G. B., and Liu, H. C. (Bot. Bull. Acad. Sin. 1999. 40: 275-281) wherein over-expression and characterization of cytosolic Cu/Zn SOD from rice in *E. coli* was achieved.

Reference may be made to document (4) by Hartman, J. R., Geller, T., Yavin, Z., Bartfeld, D., Kanner, D., Aviv, H., and Gorecki, M. (Proc. Natl. Acad. Sci. USA. 1986. 83: 7142-7146) wherein high-level expression of enzymatically active human Cu/Zn SOD was reported in *E. coli*.

Reference may be made to document (5) by Ken, C. F., Lin, C. T., Shaw, J. F., and Wu, J. L. (Marine Biotech. 2003. 5: 167-173) wherein the Cu/Zn SOD from zebrafish was over-expressed in *E. coli* and the active enzyme was purified.

Reference may be made to document (6) by Kim, T. S., Jung, Y., Na, B. K., Kim, K. S., and Chung, P. R. (Infect. Immun. 2000. 68: 3941-3948) wherein the Cu/Zn SOD gene from *Faciola hepatica* was cloned and expressed in *E. coli*.

The drawbacks are:

1. There is no SOD gene that is isolated from *Potentilla*, a source of Cu/ZnSOD that is autoclavable and functions at sub-zero temperature.
2. There is no SOD gene that is isolated from *Potentilla* and made to express in *E. coli*.
3. There is no SOD gene that is made to express in *E. coli* leading to SOD protein that is shown to be autoclavable.
4. There is no SOD gene that is made to express in *E. coli* leading to SOD protein that is shown to function at sub-zero temperature.

Comparative Data of Present SOD with other Known SOD

Present invention	Prior art
The maximum thermostability of SOD described so far is at 80° C.	The maximum thermostability of SOD is 37° C. to 50° C. reference from Bueno P., Verla, J., Gallego, G. G., and Rio del A. L. (Plant Physiol. 1995. 108: 1151-1160) wherein the thermostability of Cu/Zn SOD isolated from the cotyledon of water melon has been shown, SOD activity reduced: (a) by 40% after 4 hour of incubation at 50.° C.; (b) by 50% after 15 minute of incubation at 70° C.; (c) by 80% after 60 minute of incubation

-continued

Present invention	Prior art
stability without adding an external stabilizer [the addition of hydrogen peroxide trapping agent, polyols, and sugars etc. are required to stabilise the enzyme from other sources such as germinated plant seeds	at 80° C.; and (d) by 100% after 15 minute of incubation at 100° C. Reference may be made to Document by Miyata, K., Maejima, K., and Tomoda, K. (U.S. Pat. No. 4,563,349; Jan. 7, 1986) wherein SOD has been reported from a microorganism belonging to genus <i>Serratia</i> having the thermostability characters as follows: (a) Stable at 37° C. for 60 minutes; Inactivated by 50% when incubated at 50-60° C. for 60 minutes; and Inactivated by 100% when incubated at 80° C. for 5 minutes. External stabilizer is required to enhance the stability of the product contains this enzyme. Reported SODs do not retain their activity at ambient temperature unless stabilized by the addition of polyols, sugars or any other stabilizing agent (Bresson-Rival; Delphine; Boivin; Patrick; Linden; Guy; Perrier; Erric; Humbert; Gerard; 1999; U.S. Pat. No.
Wide range of temperature functionality from sub-zero to above 50. degree. C. temperature which would immensely enhance the utility of the enzyme and its products and be safer for use for humans.	Temperature range for SOD activity has been reported between 5 to 45. degree. C. Hakam, N. and Simon, J. P. 1996. <i>Physiol. Plant.</i> 97: 209-216). However, thermostability and lower temperature for catalyzing dismutation of O.sub.2.sup.-. are not reported for the same enzyme. There is no report for autoclavable SOD.
Present enzyme is autoclavable. When SOD is to be injected in the body, a sterile composition would be needed and for that an autoclavable SOD would be an ideal one. Moreover, in reperfusion applications and storage of organs at low temperature, an autoclavable SOD would be required which can function efficiently at low temperature as well. Apart from the use of autoclaved SOD in pharmaceuticals and medical fields, sterile SOD will also be a choice in the cosmetic and food industry.	

OBJECTS OF THE INVENTION

The main object of the invention is to provide a superoxide dismutase (SOD) Superoxide dismutase (SOD) cDNA of SEQ ID No. 2 obtained from *Potentilla atrosanguinea* containing coding gene sequence of SEQ ID No. 3 which codes for a polypeptide of SEQ ID No. 1 having Superoxide dismutase enzyme activity

Another object of the present invention is to provide a set of primers useful for the amplification of Superoxide dismutase (SOD) gene coding cDNA of SEQ ID No. 3, wherein

Forward primer 5' -ATGGCAAAGGCGTGTGCTACTT-3' and;

Reverse primer 5' -TCATCCTTGAAGCCAATAATACCA-3'

Further, another object of the present invention is to provide a method of identifying and cloning of superoxide dismutase (SOD) gene of SEQ ID NO 3, which on expression

45 gives a superoxide dismutase enzyme (EC 1.15.1.1) with the characteristics disclose in U.S. Pat. No. 6,485,950.

Yet another object of the present invention is to provide a gene responsible for autoclavable superoxide dismutase from *Potentilla*.

50 Still another object of the present invention is to provide a gene responsible for autoclavable superoxide dismutase from *Potentilla* that is also functional at sub-zero temperature.

55 Still another object of the present invention is to provide a recombinant gene of SOD, which shows activity upon autoclaving and also shows activity at low temperature, in a plasmid vector leading to a new vector which carries the nucleotide sequence synthesizing the said SOD.

60 Still another object of the present invention is to transform bacterial host *E. coli* with the above said recombinant plasmid vector for expression of the SOD gene in the bacterial host.

BRIEF DESCRIPTION OF FIGURES

65 FIG. 1 represents effect of assay temperature on SOD activity. *Potentilla* SOD expressing in *E. coli* was purified and assayed before and after autoclaving at different temperatures.

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FIG. 2 represents comparison of the nucleotide sequence of the *Potentilla* Cu/Zn SOD with sequences from other plant species. Regions of complete homology are indicated with asterisks (SEQ ID NOS 24, 3 & 25-28 are disclosed respectively in order of appearance).

FIG. 3 represents comparison of the deduced amino acid sequence of the *Potentilla* Cu/Zn SOD with sequences from other plant species. Regions of complete homology are indicated with asterisks (SEQ ID NOS 29, 1 & 30-36 are disclosed respectively in order of appearance).

FIG. 4 (A) represents expression and purification of *Potentilla* SOD in *E. coli*. C, Control; I, Protein induced by IPTG; P, Purified SOD. The gel was stained by silver staining. (B). Activity staining of the gel to depict the activity of purified SOD. P, Purified SOD.

FIG. 5 represents the result of alignment of present sod gene with the sod gene of other plant species (SEQ ID NOS 37-72 are disclosed respectively in order of appearance)

FIG. 6 represents the details of Polypeptide sequence of SOD gene SEQ ID NO. 1.

FIG. 7 represents the details of full length cDNA SOD gene of SEQ ID No. 2

FIG. 8 represents the details of coding sequence of *potentilla* SOD gene of SEQ ID NO. 3

FIG. 9 represents the details of positive cDNA clone of SEQ ID No. 4

FIG. 10 represents primer Sequence (SEQ ID NO: 5) Forward Primer and Primer sequence (SEQ ID NO: 6): Reverse primer.

FIG. 11 represents Details of primers used for RACE

(a) Primer Sequence of (SEQ ID NO. 7) GSP1: Forward primer.

(b) Primer sequence of (SEQ ID NO. 8) NES1: Reverse Primer

(c) Primer Sequence of (SEQ ID NO. 9) GSP2: Forward Primer

(d) Primer sequence of (SEQ ID NO. 10) NES2: Reverse Primer.

(Also disclosed are SEQ ID NOS 11 & 12 respectively in order of appearance).

SUMMARY OF THE INVENTION

The present invention provides superoxide dismutase gene from *Potentilla atrosanguinea* and its expression in heterologous system and comprises of a construct which carries the coding nucleotide sequence of SEQ ID 3 which is responsible for synthesis of said SOD and transformed *E. coli* producing the SOD protein. This SOD protein is autoclavable and also functions at sub-zero temperature.

DETAILED DESCRIPTION OF THE INVENTION

Accordingly, the present invention provides a superoxide dismutase (SOD) cDNA of SEQ ID No. 2 obtained from *Potentilla atrosanguinea*, wherein the said cDNA comprises 856 nucleotide bases.

In an embodiment of the present invention, the said cDNA has entire coding sequence along with pre- and post-coding sequences.

The present invention also provides a superoxide dismutase (SOD) gene coding cDNA of SEQ ID No. 3, wherein the said coding cDNA comprises 459 nucleotide bases.

Further, it also provides a superoxide dismutase (SOD) polypeptide of SEQ ID No. 1, wherein the said polypeptide comprises 152 amino acids.

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In an embodiment of the present invention, the said polypeptide is autoclavable.

In another embodiment of the present invention, the said polypeptide is functional at temperature range of $<-10^{\circ}\text{C}$. to $+80^{\circ}\text{C}$.

The present invention further provides a set of primers useful for the amplification of Superoxide dismutase (SOD) gene coding cDNA of SEQ ID No. 3, wherein

(SEQ ID NO: 5)
Forward primer 5'-ATGGCAAAGGGCGTTGCTGTACTT-3'
and;

(SEQ ID NO: 6)
Reverse primer 5'-TCATCCTTGAAGGCCAATAATACCA-3'

1. Further, it provides A method of identifying and cloning of superoxide dismutase (SOD) gene of SEQ ID NO 3 which codes for a polypeptide of SEQ ID No. 1 having Superoxide dismutase enzyme activity, wherein the said method comprising the steps of:

- isolating the mRNA from leaves of *potentilla*;
- synthesizing the cDNA from mRNA as obtained from step (a);
- constructing a cDNA library of the DNA of *potentilla* followed by the cloning of the cDNA obtained from step (b) in a suitable vector preferably in bacteriophage;
- screening the said library obtained from step (c) followed by the primary, secondary and tertiary screening for identification of positive cDNA clones;
- isolating the DNA from positive cDNA clones obtained from step (d);
- amplifying the said DNA using the primers comprising:

(SEQ ID NO: 13)
Forward Primer: 5'-GTTGTAACACGACGTGCCAGT-3'

(SEQ ID NO: 14)
Reverse Primer: 5'-CACAGGAACAGCTATGACC-3';

g) amplifying the ends of cDNA obtained from step (e) through rapid amplification of cDNA ends technique (RACE) using two set of primers to get the full length desired Superoxide dismutase (SOD) DNA of SEQ ID NO. 2 wherein the said primers comprising:

(SEQ ID NO: 7)
Forward Primer: 5'-CCAGTGGATTGCTAAGCTCATGTCCA-3'

(SEQ ID NO: 8)
Reverse Primer: 5'-GTCATCAGGGTCTGCATGGACAACAAC-3'

(SEQ ID NO: 9)
Forward Primer: 5'-ATGGTTGCATGTCAACTGGACCACATT-3'

(SEQ ID NO: 10)
Reverse Primer: 5'-TTGCATGTCAACTGGACCACATTCAA-3'

g) amplifying the ends of cDNA obtained from step (e) through rapid amplification of cDNA ends technique (RACE) using different set of primers to get the full length desired Superoxide dismutase (SOD) DNA of SEQ ID NO. 2 wherein the said primers comprising:

(SEQ ID NO: 7)
Forward Primer (GSP1):
5'-CCAGTGGATTGCTAAGCTCATGTCCA-3'

-continued

Reverse Primer (NES1): (SEQ ID NO: 8)
5'-GTCATCAGGGTCTGCATGGACAACAAC-3'

Forward Primer (GSP2): (SEQ ID NO: 9)
5'-ATGGTTGCATGTCAACTGGACCACATT-3'

Reverse Primer (NES2): (SEQ ID NO: 10)
5'-TTGCATGTCAACTGGACCACATTTCAA-3'

SMART II A Oligonucleotide: (SEQ ID NO: 11)
5'AAGCAGTGGTATCAACGCAGAGTACGCGGG-3'

3'-RACE CDS Primer A (3'-CDS): (SEQ ID NO: 73)
5'AAGCAGTGGTATCAACGCAGAGTAC(T)₃₀N₁N-3'

5'-RACE CDS Primer (5'-CDS) (SEQ ID NO: 15)
5'-(T)₂₅N₁N-3'

Universel Primer Mix A (UPM): Long: (SEQ ID NO: 16)
5'TAATACGACTCACTATAGGGCAAGCAGTG
GTATCAACGCAGAGT-3'

Universel Primer Mix A (UPM): Short: (SEQ ID NO: 17)
5'-CTAATACGACTCACTATAGGGC-3'

Nested Universel Primer A (NUP): (SEQ ID NO: 23)
5'-AAGCAGTGGTATCAACGCAGAGT-3'

- h) amplifying the coding sequence of Superoxide dismutase (SOD) of SEQ ID No. 3 using a set of primers designed from start and stop codon of full length desired Superoxide dismutase (SOD) DNA of SEQ ID NO. 2 wherein the said primers have the following sequences:

Forward Primer: (SEQ ID NO: 5)
5'-ATGGCAAAGGCGTGTGTAAGT-3'

Reverse Primer: (SEQ ID NO: 6)
5'-TCATCCTGAAGGCCAATAATACCA-3'

- i) cloning the amplified product obtained from step (g) into pQE 30 expression vector followed by the transformation it into competent *E. coli* cells to get an expression construct;
j) isolating the plasmid DNA by conventional method followed by sequencing to confirm the said SOD gene.

In an embodiment of the present invention, the polyclonal antibodies were raised against the purified SOD and used for cDNA library screening synthesized from young leaf mRNA.

In another embodiment of the present invention, library was screened and positive cDNA clones were amplified by polymerase chain reaction (hereinafter called as PCR) and two PCR products were obtained. These were sequenced and approximately 85% of the gene encoding SOD was obtained.

Further, in another embodiment of the present invention, the sequences of the said cDNA clones does not have the start and end codon and smaller by 21%.

In yet another embodiment of the present invention, primers were designed based on the sequences of positive cDNA clones and the rapid amplification of cDNA ends technique (hereinafter called as RACE) was employed to amplify the SOD full length gene.

In still another embodiment of the present invention, the said SOD gene is sequenced and analyzed comprising the sequences set forth in SEQ ID No. 2.

- 5 In still another embodiment of the present invention, the said full length SOD gene contains 856 nucleotide bases.

In still another embodiment of the present invention, the said full length SOD gene has entire coding sequence along with pre- and post-coding sequences.

- 10 In still another embodiment of the present invention, a set of primers are designed based on the full length SOD gene to amplify the superoxide dismutase (SOD) gene coding cDNA of SEQ ID No. 3.

In still another embodiment of the present invention, the said coding cDNA comprises 459 nucleotide bases.

- 15 In still another embodiment of the present invention, said SOD gene is ligated into a vector to yield a recombinant plasmid which upon transformation into a suitable *E. coli* host resulted into a clone.

- 20 In still another embodiment of the present invention, the said coding sequence of SOD gene of SEQ ID No. 3 corresponding to polynucleotides encoding Superoxide dismutase (SOD) enzyme.

Further, the present invention also provides an expression construct included sequences encoding a selectable marker and a terminator sequence.

In the present invention, leaves of *Potentilla* plant growing at Kunzum Pass (altitude 4517 m; 32° 24' N; 077° 38' E) in Lahaul and Spiti district of Himachal Pradesh in Western Himalaya of India were collected and stored in liquid nitrogen. We had earlier reported in our U.S. Pat. No. 6,485,950 that the leaves of this plant has SOD that is autoclavable and functions at sub-zero temperature. Thus the gene encoding such a SOD was identified, isolated and cloned in *E. coli* by techniques well known and routinely practiced in the art. The present SOD gene was sequenced and analyzed for its sense orientation, comprising the sequences set forth in SEQ ID No; 2. The term "sense" as used herein, refers to a substantial run of RNA bases having essentially the same bases as a specific RNA sequence (e.g., mRNA). The invention also embraced polynucleotides encoding the amino acid sequences set out in SEQ ID No. 1. The invention also provided host cells, comprising a polynucleotide of the invention in a manner that permits expression of the encoded SOD polypeptide. Suitable host cells for transformation with the SOD gene of the invention include bacterial cells e.g. *E. coli*. Polynucleotides of the invention may be introduced into the host cell as a part of a circular plasmid using the well known methods for introducing DNA into the host cell and routinely practiced in the art. Host cells of the invention are a valuable source for industrial scale production of recombinant SOD.

Polyclonal antibody, in the present invention refers to an antibody produced in the normal immune system in response to an antigen consists of a number of closely related, but not identical proteins).

55 Vector, in the present invention refers to the sequence of DNA capable of accepting foreign DNA and take the form of a circular plasmid DNA that shows resistance to a given antibiotic. The gene sequence of the invention was compared with the SOD reported from other plants to figure out the uniqueness of the gene (FIG. 2). Sequences unique to the polypeptides of the invention are recognizable through sequence comparison to other known polypeptides, and can be identified through use of alignment programs routinely utilized in the art, e.g., those made available in public sequence databases (FIG. 3). This suggested that the sequence obtained were incomplete. SEQ ID No; 3 however, shared at least 80%, at least 82%, at least 83%, at least 85%,

at least 86% sequence homology with SOD genes reported from other plants. Percent sequence "homology" with respect to polynucleotides of the invention is defined herein as the percentage of nucleotide bases in the candidate sequence that are identical to the nucleotides in the SOD coding sequence after aligning the sequences, if necessary, to achieve the maximum percent sequence identity.

It is cumulative effect/combination of amino acids for the entire 100% amino acid composition that this property is observed. This entire composition provides this protein the effect that protein has this effect.

The following examples are given by way of illustration of the present invention and should not be construed to limit the scope of the present invention.

EXAMPLE-1

Raising Antibodies Against SOD in Rabbit

Polyclonal antibodies against purified protein were raised in one-year-old male rabbit (New Zealand type). Purified SOD protein (100 µg in 500 µl of potassium phosphate buffer; pH, 7.0) was emulsified in 1 ml of Freund's complete adjuvant and administered intramuscularly using disposable syringe. Complete Freund's adjuvant was obtained from Bangalore Genei, India that contained paraffin oil, mannide monooleate as an emulsifier and heat killed *Mycobacterium tuberculosis*. After 7th days of primary injection, a booster dose (1 ml, containing 60 µg of purified protein emulsified in 1 ml of incomplete adjuvant) was administered. Adjuvant (500 µl) was thoroughly emulsified with the purified enzyme (500 µl: 100 µg) to obtain a stable antigen-adjuvant emulsion by rapidly withdrawing and expelling the antigen-adjuvant mix using a 22 gauge needle fitted to a sterile syringe. Complete emulsification was tested by placing a drop of the mixture onto a still surface of distilled water. The intactness of the droplet assures complete mixing. Antigen-adjuvant mixture (800 µl) was injected in thigh muscles of rabbit using a 22 gauge needle. Blood was collected from heart of the rabbit and allowed to clot for 2 hours at room temperature. After overnight storage at 4° C., the edges of the clot were rimmed using a Pasture pipette and centrifuged at 150×g for 5 min. Supernatant was collected and centrifuged for 15 min at 350×g to remove cell debris. Sodium azide was added to a concentration of 0.025% and the serum was stored at 4° C. After second booster dose, a small amount of blood was collected to test for the presence of the antibody using Ouchterlony Double Diffusion (hereinafter known as ODD) as described by Kanematsu, S, and Asada, K. (1990) Plant Cell Physiol. 31: 99-112. Thus, in a 85 mm petri plate, 1.5% agar prepared in 0.15 M NaCl, 20 mM potassium phosphate of pH 7.0 and 0.02% sodium azide was poured to a thickness of 3 mm. Antigen (20 µl containing 4 µg of protein) and antibody were loaded into the 3 mm diameter well cut with the help of cork-borer. Petri plate was covered and kept in a humid environment for 16-24 hour at 37° C. and examined for line of immune precipitation.

EXAMPLE-2

RNA Isolation, Quantification of RNA, Gel-electrophoresis and Purification of Poly A⁺ mRNA from Total RNA

Ribonucleic acid (hereinafter known as, RNA) from young leaf tissue of *Potentilla* was isolated using the modified guanidine hydrochloride procedure (Lal. L., Sahoo. R., Gupta. R.

K., Sharma. P. and kumar. S. Plant Molecular Biology Reporter 19: 181a-181f.). Leaf tissue (500 mg) was ground in liquid nitrogen to fine powder. Powder was transferred into a new mortar containing 5 ml of the GH buffer (8M guanidine hydrochloride, 20 mM EDTA, 20 mM MES, 100 mM βME) and was ground further. Resulting homogenate was transferred to an oak-ridge tube containing equal volume of phenol:chloroform:isoamylalcohol (25:24:1). Phases were emulsified by vortexing and separated by centrifugation at 10,000 rpm for 20 min (7° C.). Upper aqueous phase was transferred to a fresh oak-ridge tube and extracted with the equal volume of chloroform:isoamylalcohol (24:1). Resulting upper aqueous phase was transferred to a corex tube and RNA was precipitated by adding 0.2 volume of 1 M acetic acid and 0.7 volume of chilled ethanol. The tubes were kept at -72° C. for 3 h. Precipitate was pelleted by centrifugation at 10,000 rpm for 10 min at 4° C. Pellet was washed thrice using 5 ml of 3 M sodium acetate (pH 5.2) followed by final washing with 70% chilled ethanol. Pellet was dried and dissolved in minimum volume of DEPC-treated autoclaved water. RNA was quantified by measuring absorbance at 260 nm and the purity was monitored by calculating the ratio of absorbance measured at 260 and 280 nm. A value >1.8 at 260/280 nm was considered ideal for the purity of RNA used in the present investigation. The formula used to calculate RNA concentration and yield was as follows:

$$\text{Concentration of RNA } (\mu\text{g/ml}) = A_{260} \text{ (absorbance at 260 nm)} \times 40 \times \text{dilution factor}$$

$$\text{Total yield } (\mu\text{g}) = \text{concentration} \times \text{volume of stock RNA sample}$$

To check the integrity of RNA, 5-6 µg of RNA in 4.5 µl of DEPC treated autoclaved water was diluted with 15.5 µl of M1 solution (2 µl of 5×MOPS buffer, 3.5 µl of formaldehyde, and 10 µl of formamide [5×MOPS buffer: 300 mM sodium acetate, 10 mM MOPS (3-[N-morpholino]propanesulfonic acid), 0.5 mM ethylene diamine tetra-acetic acid (EDTA)] and incubated for 15 minutes at 65° C. RNA was loaded onto 1.0% formaldehyde agarose-gel after adding 2 µl of formaldehyde-gel loading buffer [50% glycerol, 1 mM EDTA (pH, 8.0), 0.25% bromophenol blue, 0.25% xylene cyanol FF], and electrophoresed at 72 volts in 1×MOPS buffer (60 mM sodium acetate, 2 mM MOPS, 0.1 mM EDTA), (Sambrook, J., Fritsch, E. F. and Maniatis, T. 1989. Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Plainview, N.Y.).

Poly-A mRNA was purified from the total RNA using dC₁₀T₃₀ oligonucleotides attached covalently to polystyrene-latex particles (Oligotex™, Qiagen Inc). Oligotex selectively binds mRNA with poly-A tail to allow purification leaving all other RNA species that lack poly-A tail. 1 mg of total RNA was used as the starting material for the isolation of the mRNA and manufacturer's instructions were followed during the procedure.

EXAMPLE-3

Construction of a Directional Complementary DNA Library (Hereinafter Referred to cDNA Library)

Poly-A⁺mRNA was used to synthesize cDNA using Time-Saver™ cDNA synthesis kit (Amersham Pharmacia Biotech. USA). First strand was synthesized using MMLV-reverse transcriptase in the presence of a bifunctional primer [5'd (AAC TGG AAG AAT TCG CGG CCG CAG GAA T₁₈)p 3](SEQ ID NO: 18) having an oligo (dT₁₈) (SEQ ID NO: 19)

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tract at the 3'-end of a restriction site for Not I. Second strand synthesis is initiated by DNA polymerase I after RNase H has nicked the RNA strand of the RNA:cDNA hybrid. The cDNA produced is extracted with phenol/chloroform and purified on a Sepharose CL-4B spun column. An Eco RI adaptor (5'-d [AATTCGGCACGAGG]-3' (SEQ ID NO: 20), [GCCGT-GCTCC]p-5') (SEQ ID NO: 21) is ligated to other end of the cDNA. cDNA was digested with Not I to release site on oligo (dT₁₈-Not I) (SEQ ID NO: 19 primer. cDNA's with Eco RI and Not I overhangs were phosphorylated to disallow self-ligation but ligation to the dephosphorylated vector.

Bacteriophage λ vector (λ ExCell Not I/Eco RI/CIP) was selected for cloning of cDNA's with Eco RI and Not I overhang generated as above. λ ExCell is derived from a λ Charon vector engineered to contain an internal, linearized copy of pExCell. Following the construction and screening using the lawn cells (*E. coli* strain NM522), the bacteriophage containing the clone of interest were used to infect a special *E. coli* strain (NP66) that enables the in vivo release of pExCell, a circular, autonomously replicating pUC-based phagemid. In vivo excision of pExCell is accomplished by site-specific recombination between attL and attR sites that flank the phagemid within the λ ExCell DNA. NP66 carries the accessory proteins require for excision under the control of a thermo-inducible promoter. In vivo excision is accomplished by infection of NP66 with λ ExCell followed by growth at 39° C., which enables the expression of these accessory proteins.

The ligated vector and the cDNA fragments were packaged in an in vitro packaging system (Ready to Go Lambda packaging kit, Amersham Pharmacia Biotech. USA). In vitro packaging system for lambda DNA uses single lysogen, which codes for all necessary packaging proteins. The extract was prepared from *E. coli* lysogen in which the prophage carries a cos mutation. The cos mutation is a deletion in the cos site which prevents the endogenous prophage from being packaged; the exogenous recombinant DNA is, however, efficiently packaged. Packaging extracts also lack Eco K and other DNA restriction systems that recognize methylated DNA, which results in the efficient packaging of methylated and unmethylated cDNA.

EXAMPLE-4

Library Screening and Identification, Amplification and Purification of Positive Phage

Library was screened using polyclonal antibodies raised against purified SOD as probe. Immobilized antibodies were detected using chemiluminescence based detection method (ECLTM western blotting analysis system, Amersham Inc.). The library was plated by making the serial dilutions of packaging reaction in SM buffer (100 mM NaCl, 8 mM MgSO₄, 50 mM Tris-HCl and 0.01% gelatin). Autoclaved and dried nitrocellulose filter membranes fitting to the size of petri plate (82 mm) were used. Membranes were soaked in 10 mM isopropyl β -D-thiogalactopyranoside (hereinafter referred to IPTG) for 5 min, air-dried and used for screening. After 6 h incubation of the plated library or as the plaques started appearing, the plates were overlaid with IPTG-soaked nitrocellulose filters. The filters were overlaid by gently holding filters with blunt ended forceps at opposite edges and centering filter over plate, without trapping any air bubble. Filter was not moved once contact is made with the plate. The plates-filters (inverted) were incubated for another 4 h at 37° C. After incubation, the plates were marked with 18-gauge needle by puncturing asymmetrically for future alignment. Filters were removed from plates with protein side up. Posi-

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tive plaques were selected using the correct orientation of the developed X-ray, filter and the plate. After marking plaques were cored out and placed in 300 μ l of SM buffer for incubation at room temperature. After 2 h, it was centrifuged at 13,000 \times g for 10 min. The supernatant was collected in a fresh sterile tube followed by addition of 30 μ l chloroform. The amplified phage was replated for secondary and tertiary screening. After tertiary screening the positive plaques were cored for in vivo phagemid release. For primary screening 10⁵ plaque forming unit (pfu) were taken and transferred to membrane. Membranes were hybridized with polyclonal antibody and developed as ECL instruction. Three strong positive clones were obtained and further taken for secondary screening which gave 70% positive signal. A few clones were taken for tertiary screening and this time all the clones gave 100% positive signal after tertiary screening. All the positive plaques were used to release the vector pExCell containing the cloned fragment.

EXAMPLE-5

In Vivo Release of Phagemid pExcell from Selected Clones

Host cells were prepared from released strain of *E. coli* NP66 in 2 \times YT medium (2 \times YT medium: 12 g trypton, 24 g yeast extract and 5 g glycerol in 1 liter of final volume in distilled/deionized water) containing 50 μ g/ml spectinomycin, 30 μ g/ml of chloramphenicol and 0.2% maltose. The culture was grown overnight at 32° C. 5 ml of 2 \times YT containing 50 μ g/ml spectinomycin, 30 μ g/ml of chloramphenicol and 0.2% maltose was incubated with 50 μ l of the overnight culture. This culture was grown at 32° C. with shaking to an A₆₀₀ of 0.5-0.8. and cells were harvested by centrifugation at 3000 \times g. The pellet was re-suspended NZCYM broth (NZCYM broth: 10 g casein hydrolysate, 5 g yeast extract, 5 g NaCl, 1 g casamino acid and 2 g MgSO₄.7H₂O in 1 liter of final volume in distilled/deionized water) containing 50 μ g/ml spectinomycin to a final A₆₀₀ of 2.0. Cells were used within 1 h. To release the pExCell, 100 μ l of the prepared NP66 cells were placed in a 15 ml sterile glass tube and incubated at 39° C. for 20 min to allow for expression of the H is proteins required for site-specific recombination between attL and attR sites. 100 μ l of the phage SM solution was added from to the cells and incubated at 39° C. for an additional 20 min. To this NP66/phage mixture, 200 μ l of 1 M sodium citrate was added to terminate the infection of NP66 with λ ExCell and 5 ml of pre-warmed (32° C.) 2 \times YT broth containing 50 μ g/ml spectinomycin was added. The culture was incubated at 32° C. with moderate shaking for 1.5 h to yield 'released culture'. To prepare overnight cultures for subsequent isolation of pExCell DNA, 50 μ l of the released culture was incubated at 37° C. in 5 ml of LB medium (LB medium: 10 g trypton, 5 g yeast extract, 10 g sodium chloride in 1 liter of final volume in distilled/deionized water) containing 100 μ g/ml ampicillin.

EXAMPLE-6

Analysis and Sequencing of Cloned cDNA

The cultures were streaked and the colonies were randomly picked up using a pipette tip. The colony was suspended in 50 μ l of lysis buffer (colony lysis buffer: TE (Tris-Cl10 mM, 1 mM EDTA, pH 8.0) with 0.1% tween 20), boiled for 10 min

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in a water bath followed by snap cooling on ice. Plasmid released in the colony lysate was amplified using 0.2 μM of each 'forward' (5'-GTTGTA AAAACGACGGCCAGT-3') (SEQ ID NO: 22) and 'reverse' (5'-CACAGGAAACAGC-TATGACC-3') (SEQ ID NO: 14) flanking primer, 20 μM of dNTPs and 1 Units of *Thermus aquaticus* (hereinafter referred to Taq) DNA polymerase (purchased from M/S. Qiagen, Germany) in 1×PCR buffer (20 mM Tris-Cl (pH, 8.4), 50 mM KCl, 1.5 mM MgCl₂). In the present invention, dNTPs refers to deoxy nucleoside triphosphate which comprises of deoxyadenosine triphosphate (hereinafter referred to dATP), deoxyguanosine triphosphate (hereinafter referred to dGTP), deoxycytidine triphosphate (hereinafter referred to dCTP) and deoxythymidine triphosphate (hereinafter referred to dTTP). Thermocycler program consisted of 30 cycles of 94° C. for 40 sec, 52° C. for 1 min and 72° C. for 2 min. This was followed by a 5 min extension at 72° C. Amplified products were run on 1.2% agarose gel in 1×TAE buffer (TAE buffer: 0.04 M Tris-acetate, 0.002 M EDTA, pH 8.5) containing ethidium bromide (final concentration of 0.5 μg/ml) and analyzed for correct size of insert by comparing with standard DNA molecular weight marker. Plasmids were isolated using QIAGEN plasmid mini kit (Cat#12125). These were quantified, checked on 1% agarose gel and sequencing was performed using the BigDye terminator (version 3.1) cycle sequencing mix (Applied Biosystems, USA) on automated DNA sequencer (ABI Prism 310, Genetic Analyzer, Applied Biosystems, USA). Protocols were followed essentially as described by respective manufacturers. Sequencing primers used were 'forward' 5'-GTTGTA AAAACGACGGC-CAGT-3' (SEQ ID NO: 22) and 'reverse' 5'-CACAGGAAA-CAGCTATGACC-3' (SEQ ID NO: 14).

INFORMATION FOR SEQ ID NO: 4

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 365 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: circular

(ii) MOLECULE TYPE: cDNA

(iii) SEQUENCE DESCRIPTION:

SEQ ID NO: 4

5' CAAGAGGGAGATGGCCAACTACTGTGACCGGAAACATTTCTGGCCTC
AAGCCTGGGCTTCATGGTTTCCATGTTTCATGCTCTTGGGGACACAACCAA
TGGTTGCATGTCAACTGGACCACATTTCAATCTGCTGGCAAAGAGCATG
GGTCTCTGAAGATGAGACTCGTCATGCTGGTGATCTTGAAATATCACT
GTTGGGGATGACGGAAGTCTGCTTCCACAATTTGACAAACAGATTC
TCTCACTGGACCACACTCTATCATTGGTAGGGCTGTTGTTGCCATGCAG
ATCTGTATGACCTTGGCAAGGGTGACATGAGCTTAGCAATCCACTGGA
AATGCTGGTGGCAGGAT 3'

EXAMPLE-7

Sequence mentioned in example 6 was searched for homology in the gene databases available at URL www.ncbi.nlm.nih.gov. using Basic Local Alignment Search Tool (hereinafter called as BLAST). It was clear from the results that the

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sequence had a homology between 80-90% with the SOD sequences submitted in the databases.

EXAMPLE-8

Cloning of Full Length Gene Using Rapid Amplification of cDNA Ends (Hereinafter Referred to Race)

Rapid amplification of cDNA ends (RACE) was used to isolate full length SOD gene from *Potentilla atrosanguinea*. RACE amplifies DNA sequences from a messenger RNA template between a defined internal site and unknown sequences of either the 3' or 5' end (Frohman, M. A., Dush, M. K. and Martin, G. R. (1988) Proc. Natl. Acad. Sci. USA 85: 8998-9002; U.S. Pat. Nos. 5,962,271 and 5962272). A set of gene specific primers were used to generate 5' and 3' ends of the gene separately. The partial cDNA sequence (SEQ ID No; 1) was used to design two sets of primers. Primers were designed such that the amplified 5' and 3' ends overlap each other over a small stretch of nucleotides. For 5' RACE, a gene specific primer (hereinafter referred to GSP1), 5'-CCAGTGGATTGCTAAGCTCATGTCCA-3' (SEQ ID NO: 7) for primary PCR and one nested gene specific primer (hereinafter referred to NES1), 5'-GTCATCAGGGTCTGCATGGACAACAAC-3' (SEQ ID NO: 8) for secondary PCR (RACE). It has been used 1 μl of 10 μM nested primers stock for secondary PCR. For 3' RACE a gene specific primer (hereinafter referred to GSP2), 5'-ATGGTTGCATGTCAACTG-GACCACATT-3' (SEQ ID NO: 9) for primary PCR and one nested primer (hereinafter referred to NES2), 5'-TTGCATGTCAACTGGACCACATTTCAA-3' (SEQ ID NO: 10) were designed. Primers were designed such that the amplified 5' and 3' ends overlap each other over a small stretch of nucleotides.

The cDNA for 5'-RACE was synthesized using a modified lock-docking oligo(dT) primer and SMART II A oligo (dT) primer. The modified oligo (dT) primer, termed the 5'-RACE CDS Primer (5'-CDS) has two degenerate nucleotide positions at the 3' end.

1 μg of total RNA was reverse transcribed in separate reactions to yield 5' and 3' RACE ready cDNA using an enzyme known as reverse transcriptase. For 5' cDNA synthesis, the reaction was carried out using 1 μM of 5'-CDS primer in a reaction mixture containing RNA and 1 μM SMART II oligo (dT) primer. The 3'-RACE cDNA is synthesized using a traditional reverse transcription procedure, but with a special oligo (dT) primer. This 3'-RACE CDS Primer A (3'-CDS) primer includes the lock-docking nucleotide positions as in the 5'-CDS and also has a portion of the smart sequence at its 5' end. Sterile H₂O was added to a final volume of 5 μl for each reaction, mixed and centrifuged. The reaction mix was incubated at 70° C. for 2 min and cooled on ice for 2 min. First-strand buffer (50 mM Tris-Cl (pH, 8.3), 75 mM KCl and 6 mM MgCl₂), 1 mM dNTPs, 2 mM DTT and reverse transcriptase were added to each reaction and incubated at 42° C. for 1.5 hr in an air incubator. Diluted the first-strand reaction product with 100 μl of Tricine-EDTA buffer (10 mM Tricine-KOH (pH 8.5), 1.0 mM EDTA) and heated tubes at 72° C. for 7 min. (Reverse transcription system was a component of SMART RACE cDNA amplification kit from BD Biosciences, USA).

Sequences of Primers used for RACE were as Follows (Purchased from BD Biosciences, USA as a Part of RACE Kit).

Primer	Primer Sequence	
SMART II A Oligonucleotide	5' - AAGCAGTGGTATCAACGCAGAGTACGCGGG - 3'	(SEQ ID NO: 11)
3' - RACE CDS Primer A (3' - CDS)	5' - AAGCAGTGGTATCAACGCAGAGTAC (T) ₃₀ N ₁ N - 3'	(SEQ ID NO: 73)
5' - RACE CDS Primer (5' - CDS)	5' - (T) ₂₅ N ₁ N - 3'	(SEQ ID NO: 15)
10X Universal Primer Mix A (UPM)	Long: 5' - TAATACGACTCACTATAGGGCAAGCAGTGGTATCAACGCAGAGT - 3'	(SEQ ID NO: 16)
	Short: 5' - CTAATACGACTCACTATAGGGC - 3'	(SEQ ID NO: 17)
Nested Universal Primer A (NUP)	5' - AAGCAGTGGTATCAACGCAGAGT - 3'	(SEQ ID NO: 23)

5' and 3' RACE cDNA were amplified using 0.2 μM GSP1, GSP2 primer and 1× universal primer (UPM), 0.2 mM dNTP and 1×BD polymerase mix. Thermocycler program consisted of 30 cycles of 94° C. for 30 sec, 68° C. for 30 sec and 72° C. for 3 min. The reaction was up-scaled to 50 μl and after the completion of PCR, 45 μl of PCR sample was run on 1.2% agarose gel in TAE buffer containing ethidium bromide (final concentration of 0.5 μg/ml) Rest of the amplified product was stored at -20° C. for secondary PCR if needed. Amplicons were cut from the gel and DNA was eluted from the gel using QIAEX II gel extraction kit from M/S Qiagen, Germany following the manufacturer's instructions. The purified DNA was cloned in pGEM-T easy vector (Promega, USA), plasmids were isolated using the Qiagen plasmid mini-isolation kit, and sequencing was performed using the BigDye terminator (version 3.1) cycle sequencing mix (Applied Biosystems, USA) on an automated DNA sequencer (ABI Prism 310, Genetic Analyzer, Applied Biosystems). Protocols were followed essentially as described by respective manufacturers. The RACE products were analyzed by BLAST.

(3) INFORMATION FOR SEQ ID NO:2

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 856 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: circular

(ii) MOLECULE TYPE: cDNA

(iii) SEQUENCE DESCRIPTION:

SEQ ID NO: 2

5' ACGGGGGGGGACTGAAATAAATAGAGAGGGTCATAGTCACATTTGCA
 TTTAGGTATCTGATTCCATTCACAAACCTCCAACCTCCACCTCTCTCTCT
 ATTTCTCTTCATCTTCATCATCTTAGGGTGCAGTGCAGATCACTTTGAAAC
 ATGGCAAAGGGCGTTGCTGTACTTAGCTCCAGTGAGGGTGTGTGGAAC
 TATCCTCTTTACCCAAGAGGGAGATGGCCCAACTACTGTGACCGGAAACA
 TTTCTGGCCTCAAGCCTGGGCTTCATGGTTTCCATGTTTCATGCTCTTGGG
 GACACAACCAATGGTTGCATGTCAACTGGACCACATTTCAATCCTGCTGG
 CAAAGAGCATGGGTCTCCTGAAGATGAGACTCGTCATGCTGGTGATCTTG
 GAAATATCACTGTTGGGGATGACGGAACTGCTTGCTTCAATGTTGAC

-continued

25 AAACAGATTCCTCTCACTGGACCACACTCTATCATTGGTAGGGCTGTTGT
 TGTCCATGCAGATCCTGATGACCTTGGCAAGGGTGGACATGAGCTTAGCA
 AATCCACTGGAAATGCTGGTGGCAGGATAGCTTGTGGTATTATTGGCCTT
 CAAGGATGAACTGGACCAGGGAGCGAAACACAGGCATCTTGTGAATTAA
 30 AACTTGAGATATTAGCGAACTCTTCGGAATTGAGTATTGAAACAAGGAAT
 ACATTGTGCATTACCAATACGTTTGGCTTAGACCTGTATTCTGTATCTCA
 ATAGTTTTCTGTGTGGTGTGTTGACAGTTATTTGTGCTCAGGCATTTTCA
 35 AAGGGATAAACACAGTAACTTTCTTGCTTTGACAAAAA
 AAAAAAA 3'

EXAMPLE-9

Amplification of Coding Sequence (Hereinafter Known as CDS) and Cloning into an Expression Vector

45 CDS of SOD was amplified by PCR using the forward primer 5'-ATGGCAAAGGGCGTTGCTGTACTT-3' (SEQ ID NO: 5) and reverse primer 5'-TCATCCTTGAAGGC-CAATAATACCA-3' (SEQ ID NO: 6) designed from start codon and stop codon. The amplified product was cloned into pQE 30 expression vector and transformed into competent *E. coli* cells. The plasmid was isolated using the standard plasmid isolation protocol (Sambrook, J., Fritsch, E. F. and Maniatis, T. 1989. Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Plainview, N.Y.) and sequencing was performed using the BigDye terminator (version 3.1) cycle sequencing mix (Applied Biosystems, USA) on an automated DNA sequencer (ABI Prism 310, Genetic Analyzer, Applied Biosystems) to confirm cloning of insert.
 60 Protocols were followed essentially as described by the manufacturer.

(4) INFORMATION FOR SEQ ID NO:3

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 459 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: circular

17

- (ii) MOLECULE TYPE: cDNA
(iii) SEQUENCE DESCRIPTION:

SEQ ID NO: 3

5' ATGGCAAAGGGCGTTGCTGTACTTAGCTCCAGTGAGGGTGTGCTGGA
ACTATCTCTTTACCCAAGAGGGAGATGGCCAACTACTGTGACCGGAAA
CATTTCTGGCCTCAAGCCTGGGCTTCATGGTTCCATGTTTCATGCTCTTG
GGGACACAACCAATGGTTGCATGTCAACTGGACCACATTTCAATCCTGCT
GGCAAAGAGCATGGGTCCTCTGAAGATGAGACTCGTCATGCTGGTGATCT
TGGAATATCACTGTTGGGATGACGGAACGCTTGCTTCACAATTGTTG
ACAAACAGATTCTCTCACTGGACCACACTCTATCATTTGGTAGGGCTGTT
GTTGTCCATGCAGATCTGTAGACCTTGGCAAGGGTGGACATGAGCTTAG
CAAATCCACTGGAAATGCTGGTGGCAGGATAGCTTGTGGTATTATTGGCC
TTCAAGGATGA 3'

(5) INFORMATION FOR Pro SEQ ID NO:1

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 152 amino acids

(B) TYPE: amino acid

(ii) MOLECULE TYPE: polypeptide

(iii) SEQUENCE DESCRIPTION:

SEQ ID NO: 1

MAKGVAVLSSSEGVAGTILFTQEGDGPPTVTGNISGLKPLHGFHVHALG
DTTNGCMSTGPHFNPAGKEHGSPEDETRHAGDLGNI TVGDDGTACTFTVD
KQIPLTGPHSIIGRAVVHADPDDLKGGHELKSKSTGNAGGRIACGI IGL
QG

EXAMPLE-10

Induction and Purification of Expressed Protein

E. coli containing SOD gene from *Potentilla* was grown at 37° C. in 100 ml of LB medium with 100 µg ml⁻¹ and 25 µg ml⁻¹ kanamycin. When that culture had grown to an absorbance of 0.6 at 600 nm, IPTG was added to a final concentration of 1 mM. CuSO₄ and ZnSO₄ were added to a final concentration of 100 ppm and 2 ppm, respectively. After inducing the expression of the SOD protein for 5 h at 37° C., cells were harvested, washed and resuspended in 4 ml of lysis buffer (50 mM NaH₂PO₄ buffer, pH 8.0, containing 300 mM NaCl and 10 mM imidazole). The cell suspension was sonicated, and the lysate was cleared by centrifugation at 12000 g and 4° C. for 20 min. The supernatant was then poured into the column loaded with nickel-nitrilotriacetic acid (Ni-NTA) agarose, washed with wash buffer (50 mM NaH₂PO₄ buffer, pH 8.0, containing 300 mM NaCl and 20 mM imidazole), and SOD protein was eluted with elution buffer (50 mM NaH₂PO₄ buffer, pH 8.0, containing 300 mM NaCl and 250 mM imidazole). The purified SOD was evaluated by 10% SDS-PAGE using silver staining to visualize the protein (FIG. 4A).

The protein estimation was performed, before and after autoclaving purified SOD that shows ±25% loss of protein. Since 50 µl of protein sample was used for assaying SOD activity, the loss in protein was calculated while calculating the enzyme activity. Reaction medium contained 0.05 M potassium phosphate buffer (pH, 7.0), 5.7×10⁻⁵ M nitroblue

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tetrazolium (hereinafter referred to NBT), 9.9×10⁻³ M methionine, 1.17×10⁻⁶ M riboflavin and 0.025% Triton X-100 in a total volume of 3.0 ml. Reaction (performed in a 30 ml glass vial) was initiated by illuminating the reaction with light intensity of 1000µ Einstein/m²/ second using a fiber optic light source (Nikon). The reaction was terminated after 2 min and the absorbance was read at 560 nm.

A control reaction was always performed wherein all the steps and components were exactly the same as described above except that purified enzyme was replaced with equal volume of homogenizing buffer. Activity of SOD is expressed as percent inhibition in color development as compared to the control reaction (higher the inhibition, higher the SOD activity). Activity data was shown in FIG. 1.

EXAMPLE-11

SOD Activity at Different Temperatures in Purified SOD

The purified SOD enzyme was assayed at temperatures ranging between -10 to 80° C. in the buffer composition as described in Example 10 except that 50% glycerol was added in the reaction mixture to avoid freezing at low temperature.

A glass beaker of 100 ml capacity was filled with either alcohol (for working at temperatures of -10, -5, 0° C.) or distilled water (for working at rest of the temperatures) or used to maintain the temperature of the reaction medium while assaying SOD. Reaction medium along with the enzyme was pre-equilibrated at desired temperature to avoid time lag in attaining the required temperature. As can be seen from FIG. 1 that the enzyme showed highest activity (87.5% inhibition) at 0° C. The enzyme was functional even up to -10° C. (82% inhibition). The enzyme is expected at temperature lower than -10° C.

Control reactions, as mentioned in Example 10, were always performed at all the temperatures.

EXAMPLE-12

Localization of SOD by Activity Staining of Native Gel

The purified SOD was localized on 10% polyacrylamide gel by activity staining as described by Beauchamp and Fridovich (Anal. Biochem. 1971; 44: 246-287). After electrophoresis, the gel was rinsed with distilled water followed by 30 min incubation in 50 ml phosphate buffer (50 mM; pH 7.0) containing 2.5 mM NBT in dark at room temperature. Gel was then immersed in 1.17×10⁻⁶ M riboflavin for 20 min, followed by exposure to light source (Nikon). Light exposure led to photogeneration of O₂⁻, which converts NBT into insoluble purple colored formazan. The purple color was developed throughout the gel except for the location where SOD was localized as shown in FIG. 4B.

Advantages:

The main advantages of the present invention are:

1. SOD gene has been cloned from *Potentilla* that is autoclavable and functions at sub-zero temperature.
2. SOD gene that is isolated from *Potentilla* has been made to express in *E. coli*.
3. SOD gene from *Potentilla* that is made to express in *E. coli* leading to synthesis of SOD protein that is autoclavable.
4. SOD gene from *Potentilla* that is made to express in *E. coli* leading to synthesis of SOD protein that is autoclavable, also functions at sub-zero temperature.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 73

<210> SEQ ID NO 1

<211> LENGTH: 152

<212> TYPE: PRT

<213> ORGANISM: *Potentilla atrosanguinea*

<400> SEQUENCE: 1

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           20           25           30
Asn Ile Ser Gly Leu Lys Pro Gly Leu His Gly Phe His Val His Ala
           35           40           45
Leu Gly Asp Thr Thr Asn Gly Cys Met Ser Thr Gly Pro His Phe Asn
           50           55           60
Pro Ala Gly Lys Glu His Gly Ser Pro Glu Asp Glu Thr Arg His Ala
           65           70           75           80
Gly Asp Leu Gly Asn Ile Thr Val Gly Asp Asp Gly Thr Ala Cys Phe
           85           90           95
Thr Ile Val Asp Lys Gln Ile Pro Leu Thr Gly Pro His Ser Ile Ile
           100          105          110
Gly Arg Ala Val Val Val His Ala Asp Pro Asp Asp Leu Gly Lys Gly
           115          120          125
Gly His Glu Leu Ser Lys Ser Thr Gly Asn Ala Gly Gly Arg Ile Ala
           130          135          140
Cys Gly Ile Ile Gly Leu Gln Gly
145           150

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<210> SEQ ID NO 2

<211> LENGTH: 856

<212> TYPE: DNA

<213> ORGANISM: *Potentilla atrosanguinea*

<400> SEQUENCE: 2

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attocattca caaacctcca acteccacct ctctctctat ttctcttcat cttcatcadc      120
ttaggggtgca ctgagatcac tttgaaacat ggcaaagggc gttgctgtac ttagctccag      180
tgagggtggt gctggaacta tcctctttac ccaagaggga gatggcccaa ctactgtgac      240
cggaaacatt tctggcctca agcctgggct tcatggtttc catgttcatt ctcttgggga      300
cacaaccaat ggttgcatgt caactggacc acatttcaat cctgctggca aagagcatgg      360
gtctctgaa gatgagactc gtcattgctg tgatcttggg aatatcactg ttggggatga      420
cggaactgct tgcttcacaa ttggttgaaa acagattcct ctcactggac cacactctat      480
cattggtagg gctgtgtgtg tccatgcaga tcctgatgac cttggcaagg gtggacatga      540
gcttagcaaa tccactggaa atgctggtgg caggatagct tgtggtatta ttggccttca      600
aggatgaact ggaccaggga gcgaaacaca ggcatcttgt tgaattaaaa cttgagatat      660
tagogaactc ttcggaattg agtattgaaa caaggaatac atttgcatt accaatacgt      720
ttggcttaga cctgtattct gtatctcaat agttttctgt gtggttgttt gacagttatt      780
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<213> ORGANISM: Potentilla atrosanguinea

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cttcattggtt tccatgttca tgctcttggg gacacaacca atggttgcac gtcaactgga    180
ccacatttca atcctgctgg caaagagcat gggctctctg aagatgagac tcgtcatgct    240
ggtgatcttg gaaatatcac tgttggggat gacggaactg cttgcttcac aattgttgac    300
aaacagattc ctctcactgg accacactct atcattggta gggctgttgt tgtccatgca    360
gatcctgatg accttgggca ggggtggacat gagcttagca aatccactgg aatgctggt    420
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<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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catttcaate ctgctggcaa agagcatggg tctcctgaag atgagactcg tcatgctggt    180
gatcttggaa atatcactgt tggggatgac ggaactgctt gcttcacaat tgttgacaaa    240
cagattcctc tcaactggacc acactctatc attggtaggg ctggttgtgt ccatgcagat    300
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aggat                                             365

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<220> FEATURE:
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

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 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

<400> SEQUENCE: 8

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<210> SEQ ID NO 9
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 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

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 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic primer

<400> SEQUENCE: 10

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<210> SEQ ID NO 11
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<400> SEQUENCE: 11

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<210> SEQ ID NO 12
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 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic oligonucleotide
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 <221> NAME/KEY: modified_base
 <222> LOCATION: (27)..(28)
 <223> OTHER INFORMATION: a, c, g, t, unknown or other

<400> SEQUENCE: 12

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<212> TYPE: DNA
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<210> SEQ ID NO 14
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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<400> SEQUENCE: 14

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<210> SEQ ID NO 15
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<220> FEATURE:
<221> NAME/KEY: modified_base
<222> LOCATION: (26)..(27)
<223> OTHER INFORMATION: a, c, g, t, unknown or other

<400> SEQUENCE: 15

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<210> SEQ ID NO 16
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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<210> SEQ ID NO 17
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<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligonucleotide

<400> SEQUENCE: 17

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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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 <400> SEQUENCE: 19

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<210> SEQ ID NO 20
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 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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 <400> SEQUENCE: 20

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<210> SEQ ID NO 21
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 <212> TYPE: DNA
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 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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 <400> SEQUENCE: 21

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 <213> ORGANISM: Artificial Sequence
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 <400> SEQUENCE: 22

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 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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 <400> SEQUENCE: 23

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<210> SEQ ID NO 24
 <211> LENGTH: 459
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 <213> ORGANISM: Malus sp.

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aagcagattc ctctgctgg accacactct atcattggta gggcggttgt tgtccacgca 360
gacctgatg accttggcaa ggggtggacat gagcttagca aatccacagg aaatgctggt 420
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<210> SEQ ID NO 25
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<213> ORGANISM: Populus sp.

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<400> SEQUENCE: 25

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acccaagaag gagatggccc aactactgta attggaaacc tttctggtct taagccaggc 120
cttcattggt tccacgtcca tgccttggga gacaccacaa atggctgcat gtcaactggg 180
ccgcatttta atcctgtagg caaggagcat ggtgcccctg aggatgagaa tgcctatgct 240
ggtgatcttg gaaatgtcac tgttggtgat gatggcactg ctgctttcac aatcattgac 300
aaacagattc ctcttactgg accacattcc attattggtt gggctggtgt tgttcatgga 360
gatcctgatg atcttggcaa gggaggacat gaactcagca aaaccactgg taatgctggc 420
ggcagagtag catgcggtat tattggtctg caaggttga 459

```

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<210> SEQ ID NO 26
<211> LENGTH: 459
<212> TYPE: DNA
<213> ORGANISM: Zea sp.

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<400> SEQUENCE: 26

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atggtgaagg ctgtggcagt tcttagtaac agtaacgaag tctcgggtac tattaacttc 60
agtcaggagg gaaatggtcc aaccactgta actggaactc ttgctggtct taagcctggc 120
ctccacggct tccatatcca tgccttggga gacaccacaa acggttgcac ttcaactgga 180
ccacatttca atcctaattg gaaggaacat ggtgcccctg aggatgagac tagacatgct 240
ggtgatcttg gaaatatcaa tgttggtgat gatggaactg taagcttcac cactactgac 300
aaccatatcc ctctcactgg aacaaactcc atcataggaa gggctggtgt tgtccatgcc 360
gatcctgatg atcttgggaa aggtggtcac gagcttagca aaactactgg aaatgctggt 420
ggcagagtag cttgtggtat tattgggttg caaggatag 459

```

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<210> SEQ ID NO 27
<211> LENGTH: 459
<212> TYPE: DNA
<213> ORGANISM: Arabidopsis sp.

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<400> SEQUENCE: 27

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atggcgaaa gaggttgcagt tttgaacagc agtgaggggtg ttacggggac tatctttttc 60
accaggaag gcgatggtgt gaccactgtg agtggaaacag tttctggcct taagcctggt 120
cttcattggt tccatgtcca tgctcttggg gacaccacta acggttgcac gtctactggt 180
ccacatttca accccgatgg taaaacacac ggtgcccctg aggatgctaa tgcacatgct 240
ggtgatctag gaaacatcac tgttggagat gatggaactg ccaccttcac aatcactgat 300
tgccagattc ctcttactgg accaaactct attgttggtg gggctggtgt tgtccatgca 360
gacctgatg acctcggaaa gggaggccat gaactcagcc tggctactgg aaacgcaggc 420

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ggccgtggtt cttgcccgcatt cattggctctc cagggctaa 459

<210> SEQ ID NO 28
 <211> LENGTH: 459
 <212> TYPE: DNA
 <213> ORGANISM: *Oryza sp.*

<400> SEQUENCE: 28

atggtgaagg ctggtgctgt gcttgctagc agtgagggtg tcaagggcac catctttttc 60
 tcccaagagg gagatggtcc gacctctgtg acgggaagtg tctctgggct caagccaggg 120
 ctccatggat tccatgtgca cgcgctcggg gacaccacta atggctgcat gtcaactgga 180
 ccacaattca atcctactgg gaaggaacat ggggcaccac aagatgagaa ccgccatgcc 240
 ggtgatcttg gaaatataac agctggagca gatggtgttg ctaatgtcaa tgtctctgac 300
 agccagatcc cccttactgg agcacactcc atcattggcc gagctgttgt tgtccatgct 360
 gatcctgatg atcttggcaa ggggtggacat gagcttagca agaccactgg aaatgctggg 420
 ggccgagttg cttgcccgaat catcggactc cagggttag 459

<210> SEQ ID NO 29
 <211> LENGTH: 152
 <212> TYPE: PRT
 <213> ORGANISM: *Malus sp.*

<400> SEQUENCE: 29

Met Val Lys Gly Val Ala Val Leu Gly Ser Ser Glu Gly Val Lys Gly
 1 5 10 15
 Thr Ile Ser Phe Val Gln Glu Gly Asp Gly Pro Thr Thr Val Thr Gly
 20 25 30
 Ser Val Ser Gly Leu Lys Pro Gly Leu His Gly Phe His Val His Ala
 35 40 45
 Leu Gly Asp Thr Thr Asn Gly Cys Met Ser Thr Gly Pro His Phe Asn
 50 55 60
 Pro Ala Gly Lys Glu His Gly Ala Pro Glu Asp Glu Leu Arg His Ala
 65 70 75 80
 Gly Asp Leu Gly Asn Ile Thr Ala Gly Asp Asp Gly Thr Ala Thr Phe
 85 90 95
 Thr Ile Val Asp Lys Gln Ile Pro Leu Ala Gly Pro His Ser Ile Ile
 100 105 110
 Gly Arg Ala Val Val Val His Ala Asp Pro Asp Asp Leu Gly Lys Gly
 115 120 125
 Gly His Glu Leu Ser Lys Ser Thr Gly Asn Ala Gly Gly Arg Val Ala
 130 135 140
 Cys Gly Ile Ile Gly Leu Gln Gly
 145 150

<210> SEQ ID NO 30
 <211> LENGTH: 152
 <212> TYPE: PRT
 <213> ORGANISM: *Arabidopsis sp.*

<400> SEQUENCE: 30

Met Ala Lys Gly Val Ala Val Leu Asn Ser Ser Glu Gly Val Thr Gly
 1 5 10 15
 Thr Ile Phe Phe Thr Gln Glu Gly Asp Gly Val Thr Thr Val Ser Gly
 20 25 30

-continued

Thr Val Ser Gly Leu Lys Pro Gly Leu His Gly Phe His Val His Ala
 35 40 45
 Leu Gly Asp Thr Thr Asn Gly Cys Met Ser Thr Gly Pro His Phe Asn
 50 55 60
 Pro Asp Gly Lys Thr His Gly Ala Pro Glu Asp Ala Asn Arg His Ala
 65 70 75 80
 Gly Asp Leu Gly Asn Ile Thr Val Gly Asp Asp Gly Thr Ala Thr Phe
 85 90 95
 Thr Ile Thr Asp Cys Gln Ile Pro Leu Thr Gly Pro Asn Ser Ile Val
 100 105 110
 Gly Arg Ala Val Val Val His Ala Asp Pro Asp Asp Leu Gly Lys Gly
 115 120 125
 Gly His Glu Leu Ser Leu Ala Thr Gly Asn Ala Gly Gly Arg Val Ala
 130 135 140
 Cys Gly Ile Ile Gly Leu Gln Gly
 145 150

<210> SEQ ID NO 31
 <211> LENGTH: 152
 <212> TYPE: PRT
 <213> ORGANISM: Populus sp.

<400> SEQUENCE: 31

Met Val Lys Ala Val Ala Val Leu Asn Ser Ser Glu Gly Val Ser Gly
 1 5 10 15
 Thr Ile Phe Phe Thr Gln Glu Gly Asp Gly Pro Thr Thr Val Thr Gly
 20 25 30
 Asn Leu Ser Gly Leu Lys Pro Gly Leu His Gly Phe His Val His Ala
 35 40 45
 Leu Gly Asp Thr Thr Asn Gly Cys Met Ser Thr Gly Pro His Phe Asn
 50 55 60
 Pro Val Gly Lys Glu His Gly Ala Pro Glu Asp Glu Asn Arg His Ala
 65 70 75 80
 Gly Asp Leu Gly Asn Val Thr Val Gly Asp Asp Gly Thr Ala Ala Phe
 85 90 95
 Thr Ile Ile Asp Phe Gln Ile Pro Leu Thr Gly Pro His Ser Ile Ile
 100 105 110
 Gly Arg Ala Val Val Val His Gly Asp Pro Asp Asp Leu Gly Lys Gly
 115 120 125
 Gly His Glu Leu Ser Lys Thr Thr Gly Asn Ala Gly Gly Arg Val Ala
 130 135 140
 Cys Gly Ile Ile Gly Leu Gln Gly
 145 150

<210> SEQ ID NO 32
 <211> LENGTH: 152
 <212> TYPE: PRT
 <213> ORGANISM: Oryza sp.

<400> SEQUENCE: 32

Met Val Lys Ala Val Val Val Leu Gly Ser Ser Glu Ile Val Lys Gly
 1 5 10 15
 Thr Ile His Phe Val Gln Glu Gly Asp Gly Pro Thr Thr Val Thr Gly
 20 25 30
 Ser Val Ser Gly Leu Lys Pro Gly Leu His Gly Phe His Ile His Ala
 35 40 45

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Leu Gly Asp Thr Thr Asn Gly Cys Met Ser Thr Gly Pro His Tyr Asn
 50 55 60

Pro Ala Gly Lys Glu His Gly Ala Pro Glu Asp Glu Thr Arg His Ala
 65 70 75 80

Gly Asp Leu Gly Asn Val Thr Ala Gly Glu Asp Gly Val Ala Asn Ile
 85 90 95

His Val Val Asp Ser Gln Ile Pro Leu Thr Gly Pro Asn Ser Ile Ile
 100 105 110

Gly Arg Ala Val Val Val His Ala Asp Pro Asp Asp Leu Gly Lys Gly
 115 120 125

Gly His Glu Leu Ser Lys Thr Thr Gly Asn Ala Gly Gly Arg Val Ala
 130 135 140

Cys Gly Ile Ile Gly Leu Gln Gly
 145 150

<210> SEQ ID NO 33
 <211> LENGTH: 152
 <212> TYPE: PRT
 <213> ORGANISM: Zea sp.

<400> SEQUENCE: 33

Met Val Lys Ala Val Ala Val Leu Gly Ser Ser Glu Gly Val Lys Gly
 1 5 10 15

Thr Ile Phe Phe Thr Gln Glu Gly Asp Gly Pro Thr Thr Val Thr Gly
 20 25 30

Ser Val Ser Gly Leu Lys Pro Gly Leu His Gly Phe His Val His Ala
 35 40 45

Leu Gly Asp Thr Thr Asn Gly Cys Met Ser Thr Gly Pro His Tyr Asn
 50 55 60

Pro Ala Ser Lys Glu His Gly Ala Pro Glu Asp Glu Asn Arg His Ala
 65 70 75 80

Gly Asp Leu Gly Asn Val Thr Ala Gly Ala Asp Gly Val Ala Asn Ile
 85 90 95

Asn Val Thr Asp Ser Gln Ile Pro Leu Thr Gly Pro Asn Ser Ile Ile
 100 105 110

Gly Arg Ala Val Val Val His Ala Asp Pro Asp Asp Leu Gly Lys Gly
 115 120 125

Gly His Glu Leu Ser Lys Ser Thr Gly Asn Ala Gly Gly Arg Val Ala
 130 135 140

Cys Gly Ile Ile Gly Leu Gln Gly
 145 150

<210> SEQ ID NO 34
 <211> LENGTH: 152
 <212> TYPE: PRT
 <213> ORGANISM: Gossypium sp.

<400> SEQUENCE: 34

Met Val Lys Ala Val Ala Val Leu Gly Ser Asn Glu Gly Val Ser Gly
 1 5 10 15

Thr Val Phe Phe Ser Gln Glu Gly Asp Gly Pro Thr Thr Val Thr Gly
 20 25 30

Asn Leu Ser Gly Leu Lys Pro Gly Leu His Gly Phe His Val His Ala
 35 40 45

Leu Gly Asp Thr Thr Asn Gly Cys Met Ser Thr Gly Pro His Phe Asn
 50 55 60

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gatcctgatg accttggcaa ggggtggacat gagcttagca aatccactgg aaatgctggt 420
ggcaggatag cttgtggtat tattggcctt caaggatga 459

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<210> SEQ ID NO 40
<211> LENGTH: 459
<212> TYPE: DNA
<213> ORGANISM: Fagus sylvatica

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<400> SEQUENCE: 40

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atggccaagg gtgtggctgt tcttagctcg aatgagggtg tttgtggcac tatctacttt 60
gcccaagaag gagatggccc aactacagta actggaaata tttctggcct taaacctgga 120
ctccatggct tccaagtgcg tgctcttggg gacacaacaa atggttgcgcat gtcaactgga 180
ccacatttca atcctgctgg caaagagcat ggtgctcctg aggatgcaaa tcgtcatgct 240
ggtgatctgg gaaatgtcaa tgttgggtgat gatggcacag tcagtttcac aataatgac 300
aaacagattc cacttttggg tccaaattcc attatcggaa gggctgttgt tgtccatgga 360
gatccagatg atcttggcaa ggggggacat gaacttagca agagcactgg aaatgctggt 420
ggcgtatag cttgtggtat cattggcttc caaggatga 459

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<210> SEQ ID NO 41
<211> LENGTH: 400
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 41

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tctttacca agaggagat ggccaacta ctgtgaccgg aaacatttct ggcctcaagc 60
ctgggcttca tggtttccat gttcatgctc ttggggacac aaccaatggt tgcattgcaa 120
ctggaccaca tttcaatcct gctggcaaag agcatgggtc tcctgaagat gagactcgtc 180
atgctggtga tcttggaaat atcactgttg gggatgacgg aactgcttgc ttcacaattg 240
ttgacaaaca gattcctctc actggaccac actctatcat tggtagggct gttgttgtcc 300
atgcagatcc tgatgacctt ggcaagggtg gacatgagct tagcaaatcc actggaaatg 360
ctggtggcag gatagcttgt ggtattattg gccttcaagg 400

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<210> SEQ ID NO 42
<211> LENGTH: 400
<212> TYPE: DNA
<213> ORGANISM: Populus tremuloides

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<400> SEQUENCE: 42

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tctttacca agaaggagat ggccaacta ctgtaattgg aaaccttct ggtcttaagc 60
caggccttca tggcttccac gtccatgccc ttggagacac cacaatggc tgcattgcaa 120
ctgggccgca ttttaatcct gtaggcaagg agcatgggtc ccctgaggat gagaatcgtc 180
atgctggtga tctgggaaat gtcactgttg gtgatgatgg cactgctgct ttcacaatca 240
ttgacaaaca gattcctctc actggaccac attccattat tggttgggct gttgttgttc 300
atggagatcc tgatgatcct ggcaaggag gacatgaact cagcaaaacc actggtaatg 360
ctggcggcag agtagcatgc ggtattattg gtctgcaagg 400

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<210> SEQ ID NO 43
<211> LENGTH: 416
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 43

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agtgaggggtg ttgctggaac tatcctcttt acccaagagg gagatggccc aactactgtg    60
accggaaca tttctggcct caagcctggg cttcatgggt tccatgttca tgetcttggg    120
gacacaacca atggttgcat gtcaactgga ccacatttca atcctgctgg caaagagcat    180
gggtctcctg aagatgagac tcgtcatgct ggtgatcttg gaaatatcac tgttggggat    240
gacggaactg cttgcttcac aattgttgac aaacagattc ctctcactgg accacactct    300
atcattggta gggctgttgt tgtccatgca gatcctgatg accttgcaa gggtgacat    360
gagcttagca aatccactgg aaatgctggt ggcaggatag cttgtggtat tattgg    416

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<210> SEQ ID NO 44
<211> LENGTH: 416
<212> TYPE: DNA
<213> ORGANISM: Manihot esculenta

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<400> SEQUENCE: 44

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agtgaggggtg ttgctgggac aatcttcttc acccaagaag gagatgggtcc aaccaccgtc    60
actggaagtg tttctggcct taagccaggg cttcatggat tccatgttca tgccttggga    120
gacacaacaa atggttgcat gtcaactggg ccacatttca accctggtgg caaagagcat    180
ggtgcccctg aggacgacat tcgtcatgct ggtgatcttg gaaatgtcac tgetggtgat    240
gatggcactg ctagtcttcac aatcgttgac aaggatattc ctctttctgg tccgattcc    300
attgtaggaa gggcagtcgt tgttcatgca gatcctgatg atcttgaaa ggggggacat    360
gaacttagca aaaccactgg aaatgctggt ggcagggtag catgtggtgt tattgg    416

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<210> SEQ ID NO 45
<211> LENGTH: 331
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 45

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actatcctct ttaccaaga gggagatggc ccaactactg tgaccggaaa catttctggc    60
ctcaagcctg ggcttcatgg tttccatggt catgctcttg gggacacaac caatggttgc    120
atgtcaactg gaccacattt caatcctgct ggcaaagagc atgggtctcc tgaagatgag    180
actcgtcatg ctggtgatct tggaaatata actgttgggg atgacggaac tgcttgcttc    240
acaattggtg aaaaacagat tcctctcact ggaccacact ctatcattgg tagggctggt    300
gttgtccatg cagatctga tgaccttggc a                                331

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<210> SEQ ID NO 46
<211> LENGTH: 331
<212> TYPE: DNA
<213> ORGANISM: Betula pendula

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<400> SEQUENCE: 46

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```

actatccact ttaccaaga agctgatggc ccaactacag taactggaaa tatttctggc    60
cttaagcctg ggctccatgg gttccatgct catgcacttg gggacacaac aaatggttgc    120
atgtcaactg ggccacattt caatcctgct ggcaaagagc atggtgctcc tgaggatgag    180
aatcgtcatg ccggtgatct gggaaatgct accgttggty atgatggtac tgccagtctc    240
acaatagttg acaagcagat tccactttct ggaccacatt ctattattgg aagggtggt    300
gttgtccacg gggatccaga tgatcttggc a                                331

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<210> SEQ ID NO 47

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<211> LENGTH: 390
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

<400> SEQUENCE: 47

ctttacccaa gagggagatg gcccaactac tgtgaccgga aacatttctg gcctcaagcc    60
tgggcttcat ggtttccatg ttcattgctct tggggacaca accaatgggt gcatgtcaac    120
tggaccacat ttcaatcctg ctggcaaaga gcatgggtct cctgaagatg agactcgtca    180
tgctggtgat cttggaata tcaactgttg ggatgacgga actgcttctc tcacaattgt    240
tgacaaacag attcctctca ctggaccaca ctctatcatt ggtagggtctg ttgttgcca    300
tgagatcct gatgacctg gcaaggtggg acatgagctt agcaaatcca ctggaaatgc    360
tgggtgcagg atagcttctg gtattattgg    390

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<210> SEQ ID NO 48
<211> LENGTH: 390
<212> TYPE: DNA
<213> ORGANISM: Populus tremula x Populus tremuloides

<400> SEQUENCE: 48

ctttacccaa gaaggagatg gtccaactac tgtaactgga agcctctgtg gtcttaagcc    60
aggccttcat ggcttccatg ttcattgcct tggagacacc acaaatgggt gcatgtcaac    120
tggcccgcct tttaatcctg taggcaaaga gcatgggtcc cctgaggatg agaatcgtca    180
tgctggtgat ttgggaaatg tcaactgttg tgatgatggc accgctactg tctcaatcat    240
tgacaaccag attcctctca ctggacaaa ttccatcgtt ggaagggctg ttgttgcca    300
tgagatcct gatgatcttg gcaagggagg acatgaactt agcaaaagca ctggtaatgc    360
tgggtgcaga gtagcatgtg gtgttattgg    390

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<210> SEQ ID NO 49
<211> LENGTH: 446
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

<400> SEQUENCE: 49

aagggcgctt ctgtacttag ctccagttag ggtgttctg gaactatcct ctttacccaa    60
gagggagatg gcccaactac tgtgaccgga aacatttctg gcctcaagcc tgggcttcat    120
ggtttccatg ttcattgctct tggggacaca accaatgggt gcatgtcaac tggaccacat    180
ttcaatcctg ctggcaaaga gcatgggtct cctgaagatg agactcgtca tgctggtgat    240
cttggaaata tcaactgttg ggatgacgga actgcttctc tcacaattgt tgacaaacag    300
attcctctca ctggaccaca ctctatcatt ggtagggtctg ttgttgcca tgcagatcct    360
gatgaccttg gcaaggtggg acatgagctt agcaaatcca ctggaaatgc tgggtgcagg    420
atagcttctg gtattattgg ccttca    446

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<210> SEQ ID NO 50
<211> LENGTH: 446
<212> TYPE: DNA
<213> ORGANISM: Plantago major

<400> SEQUENCE: 50

aaggggtgct cagtgttag cagcagttag ggtgttagtg gcaccgtcct cttttcccaa    60
gaaggagaag gaccaccac tgtaactgga aacatttctg gccttaagcc tggacttca    120
ggcttccatg ttcattgctct tgggtgacct accaacgggt gcatgtcaac aggaccacat    180

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ttcaatccgg ctgcaaaaga gcatgggtgct cctgatgatg aggttcgcca tgctggtgac 240
cttggaatg tcacagtggg agatgatgga actgcaagtt tcaccattgt tgacaagctg 300
attccgctga ctggaccaca ttccatcatt ggaagggctg ttgtgtcca tgctgacccc 360
gatgatttgg gaaggggtgg acatgaaactc agcaaaacta ccggaaatgc tggtggaaga 420
gttgcttgtg gtatcattgg tcttca 446

```

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<210> SEQ ID NO 51
<211> LENGTH: 401
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 51

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```

ggaactatcc tctttaccca agagggagat ggccaacta ctgtgaccgg aaacatttct 60
ggcctcaagc ctgggcttca tggtttccat gttcatgctc ttggggacac aaccaatggt 120
tgcatgtcaa ctggaccaca tttcaatcct gctggcaaag agcatgggtc tcctgaagat 180
gagactcgtc atgctggtga tcttgaaat atcactgttg gggatgacgg aactgcttgc 240
ttcacaattg ttgacaaaca gattcctctc actggaccac actctatcat tggtagggct 300
gttgttgtcc atgcagatcc tgatgacctt ggcaagggtg gacatgagct tagcaaatcc 360
actggaaatg ctggtggcag gatagcttgt ggtattattg g 401

```

```

<210> SEQ ID NO 52
<211> LENGTH: 401
<212> TYPE: DNA
<213> ORGANISM: Manihot esculenta

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<400> SEQUENCE: 52

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```

ggaacaatct tctttaccca agaaggagat ggtcctacca ctgtaactgg aaacatttcc 60
ggccttaagc cagggcttca tgggttccac gtccatgcc ttggagacac acaaacgggt 120
tgcatgtcaa ctgggccaca ctttaaccct tctggcaaag atcatggtgc cctgaggat 180
gagattcgtc atgctggtga tctgggaaat gtcactgctg tgatgatgg cactgctagt 240
ttcacaatta ttgacaagca tattcctctt tctggtcaaa attcaatcat aggaagggca 300
gttgttgttc atgcagatcc tgatgatctt ggcaggggag gacatgaact cagtaaaacc 360
accgaaatg ctggtggcag agtagcatgc ggtattattg g 401

```

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<210> SEQ ID NO 53
<211> LENGTH: 398
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 53

```

```

tttacccaag agggagatgg cccaactact gtgaccgaa acatttctgg cctcaagcct 60
gggcttcatg gtttccatgt tcatgctctt ggggacacaa ccaatggttg catgtcaact 120
ggaccacatt tcaatcctgc tggcaagag catgggtctc ctgaagatga gactcgtcat 180
gctggtgate ttgaaaatat cactgttggg gatgacgaa ctgcttgctt cacaattgtt 240
gacaaacaga ttcctctcac tggaccacac tctatcattg gtagggctgt tgtgtccat 300
gcagatcctg atgaccttgg caaggggtga catgagctta gcaaatccac tggaaatget 360
ggtggcagga tagcttgtgg tattattggc cttcaagg 398

```

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<210> SEQ ID NO 54

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<211> LENGTH: 398
<212> TYPE: DNA
<213> ORGANISM: Condonopsis lanceolata

<400> SEQUENCE: 54

tttaccacaag agggagatgg cccaactaaa gttactggaa gcctttcttg ccttcaacct    60
ggacctcacg gtttccatgt tcatgccctt ggtgacacaa ccaatggttg catgtcaact    120
ggtcctcatt ataatcctgc tggaaaagaa catggtgctc cagaggacga gattcgtcat    180
gctggtgacc tcgggaatgt tacagtaggc gaagacggta ctgcaaattt caccatcgtt    240
gacaaccaga ttccactatc tggacctcat tctatcattg gaagggtgtg agttgtccat    300
gctgatcctg atgatcttgg aaagggtggc catgaactca gcaaaagcac tggaaatgct    360
ggtggcagga ttgcctgtgg tatcattgga ctgcaagg                                398

```

```

<210> SEQ ID NO 55
<211> LENGTH: 394
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

<400> SEQUENCE: 55

cccaagaggg agatggccca actactgtga cggaaacat ttctggctc aagcctgggc    60
ttcatggttt ccatgttcat gctcttgggg acacaaccaa tgggtgcatg tcaactggac    120
cacatttcaa tcctgctggc aaagagcatg ggtctcctga agatgagact cgtcatgctg    180
gtgatcttgg aaatatcact gttggggatg acggaactgc ttgcttcaca attggtgaca    240
aacagattcc tctcactgga ccacactcta tcattggtag ggctgttgtt gtccatgcag    300
atcctgatga ccttggaag ggtggacatg agcttagcaa atccactgga aatgctggtg    360
gcaggatagc ttgtggtatt attggccttc aagg                                394

```

```

<210> SEQ ID NO 56
<211> LENGTH: 394
<212> TYPE: DNA
<213> ORGANISM: Gossypium hirsutum
<220> FEATURE:
<221> NAME/KEY: modified_base
<222> LOCATION: (158)
<223> OTHER INFORMATION: a, c, g, t, unknown or other
<220> FEATURE:
<221> NAME/KEY: modified_base
<222> LOCATION: (191)
<223> OTHER INFORMATION: a, c, g, t, unknown or other
<220> FEATURE:
<221> NAME/KEY: modified_base
<222> LOCATION: (221)
<223> OTHER INFORMATION: a, c, g, t, unknown or other
<220> FEATURE:
<221> NAME/KEY: modified_base
<222> LOCATION: (251)
<223> OTHER INFORMATION: a, c, g, t, unknown or other

<400> SEQUENCE: 56

cccaagaagg agatggtcca actacogtga ctgggaacct ttctggtctt aagccgggac    60
tccatggcct ccatgttcat gcccttgggg acacaactaa cgggtgcatg tcaactggac    120
cccattttaa tcctgctggc aaagagcatg gtgctcnga agatgagaac cgccatgctg    180
gtgatctagg naatgtcact gttggtgatg atggctgtgc nagcttctcc atcaccgaca    240
aacagattcc nctcacagge ccaaactcca ttatcggaag agctgtagtt gtccatgcag    300
atcccgatga ccttggaag ggcgccatg agctcagcaa aagcacagga aatgctggcg    360
gcagagtagc ttgcggtatt attggtctgc aagg                                394

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<210> SEQ ID NO 57
<211> LENGTH: 331
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

<400> SEQUENCE: 57
ctttacccaa gagggagatg gcccaactac tgtgaccgga aacatttctg gcctcaagcc   60
tgggcttcat ggtttccatg ttcattgctct tggggacaca accaatgggt gcatgtcaac   120
tggaccacat ttcaatcctg ctggcaaaga gcatgggtct cctgaagatg agactcgtca   180
tgctggtgat cttggaata tcaactgttg ggatgacgga actgcttctc tcacaattgt   240
tgacaaacag attcctctca ctggaccaca ctctatcatt ggtagggtctg ttgttgcca   300
tgcagatcct gatgacctg gcaaggttg a                               331

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<210> SEQ ID NO 58
<211> LENGTH: 331
<212> TYPE: DNA
<213> ORGANISM: Fagus sylvatica

<400> SEQUENCE: 58
ctttgccccaa gaaggagatg gcccaactac agtaactgga aatatttctg gccttaaacc   60
tggactccat ggcttccacg tgcattgctct tggggacaca acaaatgggt gcatgtcaac   120
tggaccacat ttcaatcctg ctggcaaagg gcatggtgct cctgaggatg cgaatcgtca   180
tgctggtgat ctgggaaatg tcaatgttgg tgatgatggc acagtcagtt tcacaataat   240
tgacaaacag attccacttt gtggtccaaa ttccattatc ggaaggtctg ttgttgcca   300
tggagatcca gatgatctg gcaaggttg a                               331

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<210> SEQ ID NO 59
<211> LENGTH: 399
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

<400> SEQUENCE: 59
ctttacccaa gagggagatg gcccaactac tgtgaccgga aacatttctg gcctcaagcc   60
tgggcttcat ggtttccatg ttcattgctct tggggacaca accaatgggt gcatgtcaac   120
tggaccacat ttcaatcctg ctggcaaaga gcatgggtct cctgaagatg agactcgtca   180
tgctggtgat cttggaata tcaactgttg ggatgacgga actgcttctc tcacaattgt   240
tgacaaacag attcctctca ctggaccaca ctctatcatt ggtagggtctg ttgttgcca   300
tgcagatcct gatgacctg gcaaggttg acatgagctt agcaaatcca ctggaaatgc   360
tgggtggcagg atagcttctg gtattattgg ccttcaagg                               399

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<210> SEQ ID NO 60
<211> LENGTH: 399
<212> TYPE: DNA
<213> ORGANISM: Citrus limon

<400> SEQUENCE: 60
ctttaccceg gaaggagatg gtccaacaac tgtttcagga agcctctctg gtctcaagcc   60
tggctctcat ggattccatg ttcattgctct tgggacaca acaaatgggt gcatgtctac   120
tggacccac tttaaccctg ctggaaaaga acatggagct ccagaggatg ataatcgtca   180
tgctggtgat ttaggaaatg tcaatgttag tgatgatggt actgctactt ttacagttgt   240

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tgacaatcag attcctcttt ctggacaaa ttccattatt ggaagggtg ttgtagtcca	300
cgcatatccc gatgatcttg gcaagggcgg tcatgagctg agcaaaacca ctggaaatgc	360
tggtggcaga gtagcttgcg gcataattgg cctccaagg	399

<210> SEQ ID NO 61
 <211> LENGTH: 375
 <212> TYPE: DNA
 <213> ORGANISM: *Potentilla atrosanguinea*

<400> SEQUENCE: 61

ccaagagggga gatggcccaa ctactgtgac cggaaacatt tctggcctca agcctgggct	60
tcatggtttc catgttcctg ctcttgggga cacaaccaat ggttgcctgt caactggacc	120
acatttcaat cctgctggca aagagcatgg gtctcctgaa gatgagactc gtcctgctgg	180
tgatcttggg aatatcactg ttggggatga cggaaactgct tgcttcacaa ttgttgacaa	240
acagattcct ctcaactggac cacactctat cattggtagg gctgttgttg tccatgcaga	300
tctgatgac cttggcaagg gtggacatga gcttagcaaa tccactggaa atgctggtgg	360
caggatagct tgtgg	375

<210> SEQ ID NO 62
 <211> LENGTH: 375
 <212> TYPE: DNA
 <213> ORGANISM: *Bruguiera gymnorhiza*

<400> SEQUENCE: 62

ccaagagggga gatggcccaa ctactgtaac tggaatggt tctggcctta agtcagggtc	60
tcatggcttc catgttcctg ctcttgggga cactacaaat ggttgcctgt caactgggcc	120
gcacttcaat ccaggtagca aagagcatgg tgcccctgaa gacgagaacc gtcctgcccg	180
tgacctagga aatgtaaag ttgctggatga tggcactgca acattcacia tcaactgacaa	240
tcagattcct ctactggac ccaattccat tggtggaagg gctgttgttg tccatgctga	300
tctgatgat ctgggcaagg gagggcatga acttagcaaa agcactggaa atgctggtgg	360
cagggtagca tgtgg	375

<210> SEQ ID NO 63
 <211> LENGTH: 390
 <212> TYPE: DNA
 <213> ORGANISM: *Potentilla atrosanguinea*

<400> SEQUENCE: 63

ctttacccea gagggagatg gcccaactac tgtgaccgga aacatttctg gectcaagcc	60
tgggttctcat ggtttccatg ttcctgctct tggggacaca accaatggtt gcatgtcaac	120
tggaccacat ttcaatcctg ctggcaaaga gcatgggtct cctgaagatg agactcgtca	180
tgctggtgat cttggaata tcaactgttg ggatgacgga actgcttctc tcacaattgt	240
tgacaaacag attcctctca ctggaccaca ctctatcatt ggtagggctg ttgttgtcca	300
tgcatatcct gatgacctg gcaagggctg acatgagctt agcaaatcca ctggaaatgc	360
tggtggcagg atagcttctg gtattattgg	390

<210> SEQ ID NO 64
 <211> LENGTH: 390
 <212> TYPE: DNA
 <213> ORGANISM: *Populus alba*

<400> SEQUENCE: 64

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ctttacccaa gaaggagatg gtccaactac tgtaactgga agcctctgtg gtcttaagcc 60
aggccttcat ggcttccatg ttcattgcct tggagacacc acaaatggct gcatgtcaac 120
tggcccgcac ttaaatcctg taggcaaaga gcatggtgcc cctgaggatg agaatcgta 180
tgctggtgat ttgggaaatg tcaactgttg tgatgatggc accgctactg tctcaatcat 240
tgacaaccag attcctctta ctggacaaa ttccattggt ggaagggcag ttgttgttca 300
tgagatcct gatgatcttg gcaagggagg acatgaactt agcaaaagca ctggtaatgc 360
tggtggcaga gtagcatgtg gtgttattgg 390

```

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<210> SEQ ID NO 65
<211> LENGTH: 390
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 65

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ctttacccaa gagggagatg gcccaactac tgtgaccgga aacatttctg gcctcaagcc 60
tgggcttcat ggtttccatg ttcattgcct tggggacaca accaatgggt gcatgtcaac 120
tggaccacat ttcaatcctg ctggcaaaga gcatgggtct cctgaagatg agactcgta 180
tgctggtgat cttgaaata tcaactgttg ggatgacgga actgcttctg tcacaattgt 240
tgacaaacag attcctctca ctggaccaca ctctatcatt ggtagggctg ttgttgtcca 300
tgagatcct gatgaccttg gcaagggagg acatgagctt agcaaatcca ctggaaatgc 360
tggtggcagg atagcttgtg gtattattgg 390

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<210> SEQ ID NO 66
<211> LENGTH: 390
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 66

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ctttacccaa gaaggagatg gtccaactac tgtaactgga agcctctgtg gtcttaagcc 60
aggccttcat ggcttccatg ttcattgcct tggagacacc acaaatggct gcatgtcaac 120
tggcccgcac ttaaatcctg taggcaaaga gcatggtgcc cctgaggatg agaatcgta 180
tgctggtgat ttgggaaatg tcaactgttg tgatgatggc accgctactg tctcaatcat 240
tgacaaccag attcctctta ctggacaaa ttccattggt ggaagggcag ttgttgttca 300
tgagatcct gatgatcttg gcaagggagg acatgaactt agcaaaagca ctggtaatgc 360
tggtggcaga gtagcatgtg gtgttattgg 390

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<210> SEQ ID NO 67
<211> LENGTH: 423
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 67

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cagtgagggg gttgtgggaa ctatcctctt tacccaagag ggagatggcc caactactgt 60
gaccggaaac atttctggcc tcaagcctgg gcttcatggt ttccatgttc atgctcttgg 120
ggacacaacc aatggttgca tgtcaactgg accacatttc aatcctgctg gcaaagagca 180
tgggtctcct gaagatgaga ctgcctatgc tggatgatctt ggaaatatca ctgttgggga 240
tgacggaact gcttgcttca caattgttga caaacagatt cctctcactg gaccacactc 300
tatcattggt agggctgttg ttgtccatgc agatcctgat gaccttggca aggggtggaca 360

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tgagcttagc aaatccactg gaaatgctgg tggcaggata gcttgtggta ttattggcct 420
tca 423

```

```

<210> SEQ ID NO 68
<211> LENGTH: 423
<212> TYPE: DNA
<213> ORGANISM: Populus alba

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```

<400> SEQUENCE: 68

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```

cagtgagggt gttagtggca ccatctactt caccaggaa ggagatggtc caacaactgt 60
tactgaaac gtttctggcc ttaagcctgg accccatggc tttcatgtgc atgcccttgg 120
tgacaccacc aatggttgtt tgtcaactgg acctcacttc aatcctgctg gcaaagagca 180
tggagctcct gatgatgagg ttcgccatgc tggtagcctt gggaatgtca cagttggaga 240
agatggcact gctgctttca ctattgttga caagcagata ccacttacag gaccacattc 300
cataattgga agagctgtag ttgttcatgc tgatcctgat gatcttggaa aggggtggaca 360
tgaactgagc aaaaccactg gaaatactgg tggaaagatt gcttgtggta tcaatggcct 420
tca 423

```

```

<210> SEQ ID NO 69
<211> LENGTH: 392
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 69

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acccaagagg gagatggccc aactactgtg accggaaaca tttctggcct caagcctggg 60
cttcatgggt tccatgttca tgctcttggg gacacaacca atgggttcat gtcaactgga 120
ccacatttca atcctgctgg caaagagcat gggctcctcg aagatgagac tegtcatgct 180
ggatgatctt gaaatatcac tgttggggat gacggaaactg cttgcttcaac aattgttgac 240
aaacagattc ctctcactgg accacactct atcattggta gggctgttgt tgtccatgca 300
gatcctgatg accttggcaa ggggtggacat gagcttagca aatccactgg aaatgctggg 360
ggcaggatag cttgtgggat tattggcctt ca 392

```

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<210> SEQ ID NO 70
<211> LENGTH: 392
<212> TYPE: DNA
<213> ORGANISM: Olea europaea

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<400> SEQUENCE: 70

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```

acccaagaag gagatggctc aactactggt actggaaacc tttctggcct taagcctgga 60
cttcatggct ttcattgtca cggccttggg gacaccacca atggctgtat gtcaactgga 120
cctcatttca atcctgttgg gaaagagcat ggtgcacctg gagatgagaa ccgtcatgct 180
ggatgatctt gtaatatcac agttggcgaa gatggcaccg ctgctatcaa cattgttgac 240
aagcagatag ctcttacagg accacattcc ataattggaa gagcagtagt tgtccattca 300
gatcctgatg atcttggaa ggggtggcat gaactgagca agagcactgg aaatgctggg 360
ggaagagttg cttgtgggat cattggcctt ca 392

```

```

<210> SEQ ID NO 71
<211> LENGTH: 395
<212> TYPE: DNA
<213> ORGANISM: Potentilla atrosanguinea

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<400> SEQUENCE: 71

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atcctcttta cccaagaggg agatggccca actactgtga ccgaaacat ttctggcctc    60
aagcctgggc ttcattggtt ccattgtcat gctcttgggg acacaacca tggttgcatg    120
tcaactggac cacatttcaa tcctgctggc aaagagcatg ggtctcctga agatgagact    180
cgtcattgct gtgatcttgg aatatcact gttggggatg acggaactgc ttgcttcaca    240
attgttgaca aacagattcc tctcactgga ccacactcta tcattggtag ggctgttgtt    300
gtccatgcag atcctgatga ccttggcaag ggtggacatg agcttagcaa atccactgga    360
aatgctgggt gcaggatagc ttgtggtatt attgg                                395

```

```

<210> SEQ ID NO 72
<211> LENGTH: 395
<212> TYPE: DNA
<213> ORGANISM: Solanum tuberosum

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```

<400> SEQUENCE: 72

```

```

atcctcttca ctcaagatgg agatgctcca accacagtta atggaaatat ttctggccta    60
aaactgggac ttcattggtt ccattgtcat gcccttgggtg ataccacaaa tggctgcatg    120
tcaacaggac cacattacaa tcctgctggt aaggagcatg gtgctcctga agatgaggtg    180
cgtcattgct gtgatcttgg taacatcaca gttggagaag atggtactgc atcttttact    240
attaccgaca agcagattcc tctcactggt tcacaatcca tcattggaag agctgttgtt    300
gttcattgct atcctgatga tcttggaaaag ggaggacatg agctcagtaa aagcactgga    360
aatgctggcg gaaggattgc ttgtggtatt attgg                                395

```

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<210> SEQ ID NO 73
<211> LENGTH: 57
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligonucleotide
<220> FEATURE:
<221> NAME/KEY: modified_base
<222> LOCATION: (56)..(57)
<223> OTHER INFORMATION: a, c, g, t, unknown or other

```

```

<400> SEQUENCE: 73

```

```

aagcagtggt atcaacgcag agtacttttt ttttttttt ttttttttt ttttttn      57

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We claim:

1. An isolated Superoxide dismutase (SOD) cDNA of SEQ ID No. 2 obtained from *Potentilla atrosanguinea*, wherein said cDNA comprises 856 nucleotide bases.
2. The Superoxide dismutase (SOD) cDNA as claimed in claim 1, wherein said cDNA has the entire coding sequence along with pre- and post-coding sequences.
3. An isolated Superoxide dismutase (SOD) gene of SEQ ID No. 3, wherein said gene comprises 459 nucleotide bases.
4. An isolated set of primers useful for the amplification of a Superoxide dismutase (SOD) gene of SEQ ID No. 3, wherein

- said Forward primer comprises 5'-ATG-GCAAAGGGCGTTGCTGTACTT-3' (SEQ ID NO: 5) and;
- said Reverse primer comprises 5'-TCATCCTGAAGGC-CAATAATACCA-3' (SEQ ID NO: 6).
5. An expression construct comprising a nucleotide sequence of a superoxide dismutase (SOD) gene of SEQ ID NO 3 which codes for a polypeptide of SEQ ID No. 1 having Superoxide dismutase enzyme activity, a selectable marker and a terminator sequence.

* * * * *